

JCOB REC'D PCT/PC 19 JUN 2001

FORM PTO-1390		U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE	ATTORNEY'S DOCKET NUMBER 5585-59112
TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. § 371			U.S. APPLICATION NO. (if known, see 37 C.F.R. § 1.5) 09/868605
INTERNATIONAL APPLICATION NO. PCT/GB99/04200	INTERNATIONAL FILING DATE 17 December 1999	PRIORITY DATE CLAIMED 19 December 1998	
TITLE OF INVENTION IMPROVEMENT OF TOLERANCE TO A XENOGRRAFT			
APPLICANT(S) FOR DO/EO/US Robert Ian Lechler, Nichola Jane Rogers, Anthony Dorling			
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:			
<ol style="list-style-type: none"> 1. <input checked="" type="checkbox"/> This is a FIRST submission of items concerning a filing under 35 U.S.C. § 371. 2. <input type="checkbox"/> This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. § 371. 3. <input type="checkbox"/> This is an express request to begin national examination procedures (35 U.S.C. § 371(f) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. § 371(b) and PCT Articles 22 and 39(1). 4. <input checked="" type="checkbox"/> A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date. 5. <input checked="" type="checkbox"/> A copy of the International Application as filed (35 U.S.C. § 371(c)(2)) <ol style="list-style-type: none"> a. <input type="checkbox"/> is transmitted herewith (required only if not transmitted by the International Bureau). b. <input checked="" type="checkbox"/> has been transmitted by the International Bureau. c. <input type="checkbox"/> is not required, as the application was filed in the United States Receiving Office (RO/US). 6. <input type="checkbox"/> A translation of the International Application into English (35 U.S.C. § 371(c)(2)). 7. <input checked="" type="checkbox"/> Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. § 371(c)(3)) <ol style="list-style-type: none"> a. <input type="checkbox"/> are transmitted herewith (required only if not transmitted by the International Bureau). b. <input type="checkbox"/> have been transmitted by the International Bureau. c. <input type="checkbox"/> have not been made; however, the time limit for making such amendments has NOT expired. d. <input checked="" type="checkbox"/> have not been made and will not be made. 8. <input type="checkbox"/> A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. § 371(c)(3)). 9. <input checked="" type="checkbox"/> An oath or declaration of the inventor(s) (35 U.S.C. § 371(c)(4)). (UNSIGNED) 10. <input type="checkbox"/> A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. § 371(c)(5)). 			
Items 11. to 16. below concern document(s) or information included:			
<ol style="list-style-type: none"> 11. <input checked="" type="checkbox"/> An Information Disclosure Statement under 37 C.F.R. §§ 1.97 and 1.98. 12. <input type="checkbox"/> An assignment document for recording. A separate cover sheet in compliance with 37 C.F.R. §§ 3.28 and 3.31 and the Recordal fee of \$40.00 is included. 13. <input checked="" type="checkbox"/> A FIRST preliminary amendment. <input type="checkbox"/> A SECOND or SUBSEQUENT preliminary amendment. 14. <input type="checkbox"/> A substitute specification. 15. <input type="checkbox"/> A change of power of attorney and/or address letter. 16. <input checked="" type="checkbox"/> Other items or information: <ul style="list-style-type: none"> <input checked="" type="checkbox"/> Sequence Listing. <input checked="" type="checkbox"/> Statement in Compliance. <input checked="" type="checkbox"/> Computer readable form (diskette). <input checked="" type="checkbox"/> Copy of International Search Report with cited references (see IDS). 			



24197

09868605-091201

DATE OF DEPOSIT: June 19, 2001

JG18 Rec'd PCT/PTO 19 JUN 2001

U.S. APPLICATION NO. (If known, see 37 C.F.R. § 1.51) <div style="font-size: 1.5em; font-weight: bold;">09/868605</div>	INTERNATIONAL APPLICATION NO. PCT/GB99/04200	ATTORNEY'S DOCKET NUMBER 5585-59112
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17. ☒ The following fees are submitted:

BASIC NATIONAL FEE (37 C.F.R. §§ 1.492(a)(1)-(5)):

Neither International Preliminary Examination fee (37 C.F.R. § 1.482) nor International Search fee (37 C.F.R. § 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO..... \$1,000.00

International Preliminary Examination fee (37 C.F.R. § 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO..... \$860.00

International Preliminary Examination fee (37 C.F.R. § 1.482) not paid to USPTO but International Search fee (37 C.F.R. § 1.445(a)(2)) paid to USPTO..... \$710.00

International Preliminary Examination fee paid to USPTO (37 C.F.R. § 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4)..... \$690.00

International Preliminary Examination fee paid to USPTO (37 C.F.R. § 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4)..... \$100.00

ENTER APPROPRIATE BASIC FEE AMOUNT =		\$ 860.00
Surcharge of \$130.00 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 C.F.R. § 1.492(e)).		\$
CLAIMS	NUMBER FILED	NUMBER EXTRA
Total claims	26 - 20 =	6
Independent Claims	2 - 3 =	0
MULTIPLE DEPENDENT CLAIM(S) (if applicable)		+ \$270.00
TOTAL OF ABOVE CALCULATIONS =		\$ 968.00
<input checked="" type="checkbox"/> Reduction of 1/2 for filing by small entity. Small entity status is claimed for this application.		\$ 484.00
SUBTOTAL =		\$ 484.00
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 Months from the earliest claimed priority date (37 C.F.R. §§ 1.492(f)).		\$
TOTAL NATIONAL FEE =		\$ 484.00
Fee for recording the enclosed assignment (37 C.F.R. § 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 C.F.R. §§ 3.28, 3.31). \$40.00 per property.		\$
TOTAL FEES ENCLOSED =		\$ 484.00
		REFUND → \$
		CHARGE → \$

a. ☒ A check in the amount of \$ 484.00 to cover the above fees is enclosed.

b. ☐ Please charge my Deposit Account No. _____ in the amount of \$ _____ to cover the above fees. A duplicate copy of this sheet is enclosed.

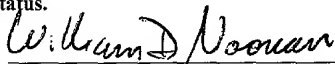
c. ☒ The Director is hereby authorized to charge any additional fees that may be required, or credit any overpayment, to Deposit Account No. 02-4550. A duplicate copy of this sheet is enclosed.

d. ☒ Please return the enclosed postcard to confirm that the items listed above have been received.

NOTE: Where an appropriate time limit under 37 C.F.R. § 1.494 or § 1.495 has not been met, a petition to revive (37 C.F.R. § 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

KLARQUIST SPARKMAN CAMPBELL
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 William D. Noonan, M.D.
 NAME
 30,878
 REGISTRATION NUMBER

cc: Docketing

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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of: Lechler

Art Unit:

Application No.

CERTIFICATE OF MAILING

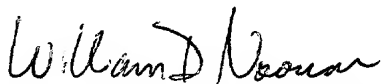
Filed: Herewith

I hereby certify that this paper and the documents referred to as being attached or enclosed herewith are being deposited with the United States Postal Service on June 19, 2001 as Express Mail No. EL828141257US in an envelope addressed to: BOX PCT, COMMISSIONER FOR PATENTS, WASHINGTON, D.C. 20231.

For: IMPROVEMENT OF TOLERANCE TO A
XENOGRAFT

Examiner:

Date: June 19, 2001



William D. Noonan, M.D., Attorney for Applicant

BOX PCT
COMMISSIONER FOR PATENTS
WASHINGTON, D.C. 20231

PRELIMINARY AMENDMENT

Before calculating the filing fee for the present application, please amend the claims as follows:

1. (Amended) A method of improving tolerance to a xenograft comprising: immunising a mammal with an immunogen comprising at least one T-cell epitope and at least one porcine polypeptide B-cell epitope, wherein said B-cell epitope is capable of mediating rejection of said xenograft.
2. (Amended) A method according to Claim 1, wherein said B-cell epitope is a peptide derived from at least one porcine polypeptide selected from the group of CD40, CD80, CD86 and VCAM.
3. (Amended) A method according to Claim 1, wherein said peptide is selected from at least one peptide represented in Figure 22.
4. (Amended) A method according to Claim 1, wherein said peptide is selected from at least one peptide represented in Figure 24.

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5. (Amended) A method according to Claim 1, wherein said peptide is selected from at least one peptide represented in Figure 26.
6. (Amended) A method according to Claim 1, wherein said T-cell epitope comprises a tetanus toxoid polypeptide.
7. (Amended) A composition comprising an immunogen characterised in that said immunogen comprises at least one B-cell epitope and at least one T-cell epitope wherein said B-cell epitope comprises a porcine epitope involved in mediating xenograft rejection.
8. (Amended) A composition according to Claim 7, wherein said porcine epitope comprises a porcine polypeptide expressed by vascular endothelial cells of said xenograft.
9. (Amended) A composition according to Claim 7, wherein said B-cell epitope is selected from the group of CD40, CD86, CD80 and VCAM.
10. (Amended) A composition according to Claim 9, wherein said B-cell comprises at least one peptide as represented in Figure 22.
11. (Amended) A composition according to Claim 9, wherein said B-cell epitope comprises at least one peptide as represented in Figure 24.
12. (Amended) A composition according to Claim 9, wherein said B-cell epitope comprises at least one peptide as represented in Figure 26.
13. (Amended) A composition according to Claim 9, wherein said B-cell epitope comprises an extracellular domain of CD86.

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14. (Amended) A composition according to Claim 7, wherein said T-cell epitope comprises a tetanus toxoid epitope.

15. (Amended) A composition according to Claim 7, wherein said composition further comprises a carrier capable of enhancing the immune response to said immunogen.

16. (Amended) An antibody, or the effective part thereof, wherein said antibody is capable of distinguishing between porcine polypeptides according to Claim 7, and the homologous polypeptides of the mammal receiving said xenograft.

17. (Amended) An antibody according to Claim 16, wherein said antibody is monoclonal.

18. (Amended) An antibody according Claim 16, wherein said antibody is a modified antibody comprising at least one detectable label.

19. (Amended) A method to monitor an immune status of a mammalian recipient of a xenograft comprising:

- i) removing a sample from a xenograft recipient to be tested;
- ii) contacting said sample to the antibody according to Claim 16; and
- iii) monitoring expression of a porcine polypeptide shown in Figures 22, 24,

or 26.

20. (Amended) A method of treating a mammal prior to receiving a xenograft, comprising:

- i) immunising a mammal with an immunogenic composition according to Claim 7;

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- ii) assessing an immune status of said mammal to said immunogenic composition;
- iii) transplanting said xenograft tissue/organ into a recipient mammal; and
- iv) monitoring a rejection response to said xenograft.

21. (Amended) A method according to Claim 20, wherein said xenograft is of porcine origin and said mammal is human.

22. (Amended) A method according to Claim 20, wherein said xenograft comprises at least one vascularised graft and/or immunogenic porcine cell/tissue.

23. (Amended) A method according to Claim 20, wherein said xenograft comprises pancreatic islets.

24. (New) The method Claim 1, wherein said B-cell epitope has less than 75% sequence identity to a corresponding region of an equivalent human polypeptide.

25. (New) The method of Claim 7, wherein said B-cell epitope has less than 75% sequence identity to a corresponding region of an equivalent human polypeptide.

26. (New) The method of Claim 16, wherein said B-cell epitope has less than 75% sequence identity to a corresponding region of an equivalent human polypeptide.

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REMARKS

The claims in this application have been amended, solely for the purpose of complying with U.S. claiming conventions.

Respectfully submitted,

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By



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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of: Lechler et al.

Art Unit:

Application No.

CERTIFICATE OF MAILING

Filed: Herewith

For: IMPROVEMENT OF TOLERANCE TO A
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I hereby certify that this paper and the documents referred to as being attached or enclosed herewith are being deposited with the United States Postal Service on June 19, 2001 as Express Mail No. EL828141257US in an envelope addressed to: BOX PCT, COMMISSIONER FOR PATENTS, WASHINGTON, D.C. 20231.


William D. Noonan, M.D., Attorney for Applicant

STATEMENT IN COMPLIANCE WITH 37 C.F.R. § 1.821(f)

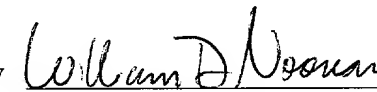
BOX PCT
COMMISSIONER FOR PATENTS
Washington, DC 20231

Sir:

In compliance with 37 C.F.R. § 1.821(f), the undersigned declares that the nucleotide and/or amino acid sequences presented in the paper copy of the "Sequence Listing" submitted herewith are the same as the sequences contained in the computer-readable form of said "Sequence Listing." No new matter has been added.

Respectfully submitted,

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09/868605-091201

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**Marked-up Version of Amended Claims
Pursuant to 37 C.F.R. §§ 1.121(b)-(c)**

CLAIMS

1. A method of improving tolerance to a xenograft comprising[;] :
immunising a mammal with an immunogen comprising at least one T-cell epitope and at least one porcine polypeptide B-cell epitope, [characterised in that] wherein said B-cell epitope is [derived from at least one porcine polypeptide involved in] capable of mediating [the] rejection of said xenograft.
2. A method according to Claim 1, [characterised in that] wherein said B-cell epitope is a peptide derived from at least one porcine polypeptide selected from[;] the group of CD40[;] , CD80[;] , CD86 [or] and VCAM.
3. A method according to Claim 1, [or 2 characterised in that] wherein said peptide is selected from at least one peptide represented in Figure 22.
4. A method according to Claim 1, [or 2 characterised in that] wherein said peptide is selected from at least one peptide represented in Figure 24.
5. A method according to Claim 1, [or 2 characterised in that] wherein said peptide is selected from at least one peptide represented in Figure 26.
6. A method according to [Claims 1 - 5 characterised in that] Claim 1, wherein said T-cell epitope [is derived from] comprises a tetanus toxoid polypeptide.
7. A composition comprising an immunogen characterised in that said immunogen [has] comprises at least one B-cell epitope and at least one T-cell epitope wherein said B-cell epitope [is derived from at least one] comprises a porcine [polypeptide] epitope involved in mediating xenograft rejection.

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8. A composition according to Claim 7₂ [characterised in that] wherein said porcine epitope comprises a porcine polypeptide [is] expressed by vascular endothelial cells of said xenograft.

9. A composition according to [Claims 7 or 8 characterised in that] Claim 7, wherein said B-cell epitope is [derived from at least one porcine polypeptide] selected from[;] the group of CD40[;] , CD86[;] , CD80[;] and VCAM.

10. A composition according to Claim 9₂ [characterised in that] wherein said B-cell epitope [is selected from] comprises at least one peptide as represented in Figure 22.

11. A composition according to Claim 9₂ [characterised in that] wherein said B-cell epitope [is selected from] comprises at least one peptide as represented in Figure 24.

12. A composition according to Claim 9₂ [characterised in that] wherein said B-cell epitope [is selected from] comprises at least one peptide as represented in Figure 26.

13. A composition according to [Claims 9 or 12 characterised in that] Claim 9, wherein said B-cell epitope [is derived from the] comprises an extracellular domain of CD86.

14. A composition according to [Claims 7 - 13 characterised in that] Claim 7, wherein said T-cell epitope [is derived from] comprises a tetanus toxoid epitope.

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15. A composition according to [Claims 7 - 14 characterised in that] Claim 7, wherein said composition further comprises a carrier capable of enhancing the immune response to said immunogen.

16. An antibody, or the effective part thereof, [characterised in that] wherein said antibody is capable of distinguishing between porcine polypeptides according to [Claims 7 – 15] Claim 7, and the homologous polypeptides of the mammal receiving said xenograft.

17. An antibody according to Claim 16, [characterised in that] wherein said antibody is monoclonal.

18. An antibody according to [Claims 16 or 17 characterised in that] Claim 16, wherein said antibody is a modified [with] antibody comprising at least one detectable label.

19. A method to monitor [the] an immune status of a mammalian recipient of a xenograft comprising:

- iii) removing a sample from a xenograft recipient to be tested;
- iv) contacting said sample to the antibody according to [Claims 16 – 18]

Claim 16; and

iii) monitoring [the] expression of [the] a porcine polypeptide [according to Claims 4 –8] shown in Figures 22, 24, or 26.

20. A method [to treat] of treating a mammal prior to receiving a xenograft, comprising:

i) immunising a mammal with an immunogenic composition according to [Claims 7 – 15] Claim 7;

ii) assessing [the] an immune status of said mammal to said immunogenic composition;

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iii) [transplantation of] transplanting said xenograft tissue/organ into a recipient mammal; and

iv) monitoring [the] a rejection response to said xenograft.

21. A method according to Claim 20, [characterised in that] wherein said xenograft is of porcine origin and said mammal is human.

22. A method according to Claim 20, [or 21 characterised in that] wherein said xenograft [is] comprises at least one vascularised graft and/or immunogenic porcine cell/tissue.

23. A method according to Claim 20, [characterised in that] wherein said xenograft [is] comprises pancreatic islets.

24. (New) The method Claim 1, wherein said B-cell epitope has less than 75% sequence identity to a corresponding region of an equivalent human polypeptide.

25. (New) The method of Claim 7, wherein said B-cell epitope has less than 75% sequence identity to a corresponding region of an equivalent human polypeptide.

26. (New) The method of Claim 16, wherein said B-cell epitope has less than 75% sequence identity to a corresponding region of an equivalent human polypeptide.

Rec'd PCT/PTO 19 JUN 2001

IMMUNOSUPPRESSION

1. FIELD OF THE INVENTION

5 This invention relates to immunosuppression and, more particularly, to immunosuppression in the context of xenotransplantation.

2. BACKGROUND TO THE INVENTION

10 Despite the established success of allogeneic organ transplantation, the increasing disparity between the supply and demand of organs must be overcome. Increasing the supply of allogeneic organs does not offer a satisfactory solution because even if all usable organs were transplanted this would still not meet the existing demand (1,2). This
15 has led to a resurgence of interest in xenotransplantation (the transplantation of organs between animals of different species) as a viable and attractive alternative.

Xenotransplantation research has recently focused on the pig as a suitable animal donor in terms of size, physiological compatibility and breeding characteristics (3,4). Until
20 recently however, discordant xenotransplantation has been limited by the inevitable occurrence of humorally-mediated hyperacute rejection (HAR) which rapidly triggers organ rejection upon revascularisation. HAR is the fate of most organs transplanted between discordant species. Recently, significant advances have been made in understanding the immunological basis of HAR, and many approaches have been
25 employed to overcome it. Of significance, a variety of transgenic strategies are currently being employed including the expression of regulators of complement activity on porcine endothelial cells (5). It is foreseeable that short-term xenograft survival will soon be achieved (6). The recent advances in overcoming HAR have highlighted subsequent immunological barriers which must be surmounted to enable long-term xenograft
30 survival. Both humoral and cellular arms of the immune response appear to play a role in the downstream events of immunological rejection. Clearly the most important of which is the existence of a formidable T cell mediated rejection response (7-11) previously obscured by the dominant role of HAR. *In vitro*, human T cells have been demonstrated

to play a central role in the recognition of xenogeneic cells (7,8,12) following sensitisation via the direct and indirect T cell activation pathways, which have been well documented for allorecognition and allograft rejection (13). Knowledge of the cellular mechanisms underlying allorecognition has provided an important basis for the investigation of the T cell mediated xenoresponse.

At present, the major therapies to prevent cell mediated rejection of organ transplants rely on systemic immunosuppressive drugs or monoclonal antibody (Mab) therapy directed against targets such as CD3, CD4, CD25, (14). Following reports that strong T cell xenoresponses can be generated *in vitro* (7,8,12), control of xenograft rejection may require levels of immunosuppression much greater than the current standard doses. Such a strategy would not be desired in a xenograft context. Drugs must be taken for life, depress the entire immune system and result in an increased risk of infection and susceptibility to cancer (14). For the applicability of xenotransplantation to the clinic, targeting graft-specific strategies for tolerance induction/immunosuppression would clearly be highly advantageous. Whilst this has been difficult to achieve in an allotransplant context, xenotransplantation offers greater potential - with differences between species providing the option for the generation of reagents that are truly graft specific. In addition, there is the opportunity for the manipulation of both the porcine donor organ, and the human recipient's immune system, prior to transplantation (1).

3. DETAILED BACKGROUND

3.1 T cell activation and proliferation

Optimal proliferation of T cells, although initiated via ligation of the antigen specific CD3/TCR complex (Signal 1) requires additional costimulatory signals (Signal 2) (15,16,17) which are usually supplied by the antigen presenting cell (APC). Whilst antigenic stimulation of T cells in the presence of signal 2 induces T cell activation and proliferation (18), exposure of T cells to MHC-antigen complexes in their absence leads to aborted T cell proliferation and the development of clonal anergy (19,20). Manipulation of APC by aldehyde fixation (20,21) or heat treatment (19) has been

demonstrated to abrogate the ability of such cells to activate alloreactive T cells, without altering levels of MHC-II surface expression. Thus T cell receptor occupancy alone is insufficient to fully activate the T cell (17). Anergic T cells are best characterised by their lack of IL-2 production and their continued inability to produce IL-2 on subsequent exposure to antigen (22). Thus, confirming the two signal model of activation as predicted by Lafferty *et al* (23). For T cells to respond to a given antigenic stimulus, multiple activation signals are required from the APC (23).

The *in vivo* induction of T cell anergy in the absence of a secondary signal was first demonstrated by Jenkins and Schwartz in 1986 (24) using chemically fixed APC to present specific peptide to CD4 T helper clones. A multitude of *in vitro* and *in vivo* data has since been produced supporting the hypothesis that signal 1 in isolation fails to activate T cells (22), and that costimulatory signalling results from contact with other cells rather than via soluble factors. Fibroblasts transfected with human Class II MHC molecules, but not expressing the appropriate CS signals (lacking signal 2) can efficiently present antigen to class II restricted CD4 T cell clones, but these fail to cause antigen specific T cell proliferation, rendering cells anergic. The context in which T cells first encounter antigen therefore has an important bearing on subsequent immune responsiveness.

Thus, costimulatory molecules are essential for T cell activation and multiplication and result from interactions between receptors on T cells and their ligands expressed on the APC. The costimulatory signal itself, however, is neither antigen specific nor MHC restricted (25). In recent years the molecular interactions involved in mediating costimulation have been well defined. The two key pathways involve (i) B7-1, B7-2 (members of the B7 family) and (ii) CD40, which are expressed on the APC, and their counter-receptors CD28 and CD40 ligand (CD40L) respectively expressed on T cells. A large body of evidence, both *in vivo* and *in vitro*, clearly defines the crucial roles played by B7-1, B7-2 and CD40 in providing T cell costimulation (26-36). Furthermore, the simultaneous blockade of signalling via CD28-B7 and CD40-CD40L in an allotransplant

context prevented the onset of allograft rejection (37,38). *In vivo*, targeting the B7/CD28 interaction has been shown to prevent T cell sensitisation to graft antigen, thereby prolonging graft survival (38,39).

5 T cells can be sensitised against xenoantigens via one of two pathways - the direct and indirect pathways, which are analogous to the well documented T cell activation pathways against alloantigens (Figure 1). Direct recognition requires that the recipient T cells recognise intact xeno MHC-molecules complexed with peptide on donor stimulator cells. In contrast, indirect recognition requires that recipient APC process the xenoantigen
10 prior to presentation to recipient T cells in the context of recipient MHC II. Self MHC II restricted T cells with specificity for the xenoantigen will recognise the peptide and respond. Whilst the majority of data reported is of indirect xenorecognition responses, cell mediated rejection via the direct route has also been documented (7,8,9,11,12,40,41,42). Vigorous human T cell proliferative responses directed against
15 porcine tissues *in vitro* have been documented from studies both in this laboratory and others.

3.2 Costimulatory molecules

The crucial role played by costimulatory molecules in determining the result of TCR-CD3
20 receptor engagement with MHC and peptides has been demonstrated extensively both *in vivo* and *in vitro*. Anti-costimulatory molecule strategies aimed at either the receptors or their ligands are being used as therapeutic strategies for altering the immune response. Such approaches have been tested in model transplant systems to alter cell mediated responses thereby preventing graft rejection (14,37,38,43-47).

25 B7-1 (B7/BB1, CD80) and B7-2 (CD86) both belong to the immunoglobulin superfamily and are heavily glycosylated transmembrane proteins (25). B7-1, a B cell activation molecule was first identified in 1989 (27), followed by B7-2 in 1993 (49). Both human B7-1 and B7-2, and the murine homologues have now been cloned and functionally
30 characterised (25). B7-1 and B7-2 are constitutively expressed on splenic and blood

dendritic cells and are induced on B cells and monocytes upon activation (34,50,). B7-1 and 2 are highly homologous and are the natural ligands for the T cell antigen CD28 (50). Cytotoxic T lymphocyte antigen-4 (CTLA-4), a cell surface glycoprotein has been identified as a second receptor for the B7 family of molecules (51) and is homologous to CD28 with 31% sequence identity. Both B7 isoforms bind to CTLA-4 with higher affinity than to CD28 (30,50,52). Whilst CD28-B7 receptor engagement results in an APC-derived costimulatory signal involved in antigen specific IL-2 production both *in vivo* and *in vitro* (53,54), CTLA4 appears to function as a negative regulator of T cell activation (55, 56, 57). Cross-linking by anti-CTLA4 antibodies has been demonstrated to antagonise CD28 ligation (58) and, in addition, CTLA4 knock-out mice die due to uncontrolled lymphocyte proliferation within the first few weeks of life (59). Thus, CTLA4 ligation is thought to be crucial for the maintenance and regulation of immune responses. The underlying mechanisms have not, however, been clearly defined.

Among costimulatory molecules, the B7 family appears to be unique, since ligation by CD28 of either B7-1 or B7-2 is both necessary and sufficient to prevent the induction of anergy (34). The CD28-B7 interaction is thought to deliver crucial signals to sustain proliferation of activated T cells. These observations are supported by *in vitro* data showing that whilst cells deficient in B7 fail to stimulate a primary MLR, transfectants expressing high levels of B7 gained the capacity to stimulate the production of IL-2 by alloreactive T cells and to co-stimulate a polyclonal population of purified T cells cultured with immobilised anti-CD3 Mab (31). Artificial APC generated by stably transfecting NIH-3T3 cells with HLA-DR7, B7 or both, clearly demonstrated that following presentation of tetanus toxoid (TT) optimal T cell proliferation and IL-2 production resulted only when both molecules were present. In the absence of B7, clonal anergy resulted (58).

Porcine B7-2 (PoB7-2) has been cloned from aortic endothelial cells (60). Following transient transfection of porcine B7-2, human umbilical vein endothelial cells strongly costimulated IL-2 production by human T cells. This costimulation of human T cells by

poB7-2 was shown to be as effective as costimulatory signals provided by human B7-1 or B7-2 and could be specifically blocked by huCTLA4Ig. Thus poB7-2 strongly contributes to the immunogenicity of porcine endothelium (60).

- 5 Although B7-1 and B7-2 mediated interactions appear to be central to the development of T cell specific immunity, additional costimulatory pathways of importance exist. The most crucial of which involves the CD40 and CD40 ligand (CD40L) interaction (34).

CD40 is a 50kDa surface glycoprotein belonging to the TNF-receptor superfamily. CD40
10 is expressed on various APC including among others, monocytes, dendritic cells and activated macrophages. Other cell types including endothelium also express CD40 (34). Its counter-receptor CD40L (CD154, gp39, TRAP) is a 33 kDa type II integral membrane protein (34,36) transiently expressed on activated CD4 T cells. The CD40-CD40L interaction has been demonstrated to play an important role in both the humoral and
15 cellular arms of the immune response with a dominant role in B cell activation. Whilst cross linking of CD40 on B cells is essential for B cell growth and isotype switching, it also results in the upregulation of B7 expression (50). Levels of B7 expression (and thus APC capacity) of monocytes and dendritic cells are clearly unregulated following CD40 signalling (34). Data from CD40 knock-out mice demonstrated that CD40L signalling
20 following ligation by CD40 plays an important role in T cell activation (61). Transfection of the murine P815 mastocytoma cells with CD40 (or B7-1) enabled previously non-stimulatory P815 cells to mediate the costimulation necessary for polyclonal T cell activation and the generation of cytokines (34). CD40-CD40L interactions have also been demonstrated to play a critical role in allograft rejection (62,63).

25

Resting B cells do not normally express B7-1/B7-2 at high levels until they are activated (50). Activation of B cells following simultaneous engagement of MHC-peptide/TCR and CD40-CD40L leads to the upregulation of B7 family members on B cells, thereby enhancing the stimulation and subsequent activation of T cells (34,36). Thus, the
30 CD40-CD40L interaction influences costimulatory activity by inducing expression of the

B7 family of molecules and perhaps other costimulatory molecules, thereby playing a key role in T cell activation . The clear synergistic effects of CD40 and B7 indicate the importance of both costimulatory pathways for the initiation and amplification of T cell dependent immune responses (38). CD40-CD40L interactions have also been shown to
5 play a crucial role in the generation of cytotoxic T lymphocyte (CTL) responses by modifying the functional status of a dendritic cell (64,65,66)

Extensive studies have demonstrated the importance of blocking B7-CD28 and/or CD40-CD40L interactions in the context of both allo and xenotransplantation. Data strongly
10 supporting this includes the use of CTLA4Ig to block signalling via CD28-B7 resulting in enhanced graft survival and the prevention of chronic rejection in a rat cardiac allograft model (44,45) and a murine aortic allograft model (43). In these models, administration of CTLA4Ig caused partial (44) or complete (46) tolerance to graft antigen by inducing T cell anergy. Treatment of allo pancreatic islet transplants with anti-B7-2 and B7-1
15 antibody has also been demonstrated to inhibit transplant rejection (14). Similar results were obtained in models inhibiting CD40 signalling in a mouse cardiac allotransplant models (37,47,62). Two studies detailing the simultaneous blockade of signalling via CD28-B7 and CD40-CD40L prevented the onset of allorejection. Concurrent prolonged inhibition of both pathways completely abrogated the onset of chronic rejection in a
20 mouse allo model (37) and in a skin and heart allo model (38).

In the realm of xenotransplantation, Lenschow and colleagues have, demonstrated long-term donor specific tolerance of human islets transplanted into mice with concomitant treatment with CTLA4Ig (46). Graft specific tolerance was demonstrated to be a direct
25 consequence of inhibiting recognition via B7 expressing APC. In addition, Tran *et al* (67) demonstrated short term suppression with CTLA4-Fc treatment. There is limited data available on the simultaneous blockade of both pathways in the xenotransplantation context, with the prolonged survival of rat and porcine skin transplanted into murine recipients (63).

In vitro and *in vivo* data have clearly demonstrated that targeting the interactions mediated by either the CD28-B7, CD40-CD40L, or both pathways has prevented the sensitisation of T cells to alloantigen and xenoantigen from engrafted tissue thereby prolonging graft survival ().

5

As noted above, T- cell mediated graft rejection is well documented. The immune system can mount alternate or additional cell mediated rejection mechanisms. These mechanisms are illustrated by the function of various molecules expressed by, *inter alia*, endothelial cells. VCAM is a cell adhesion molecule, expressed by endothelial cells, that is thought to have a role in leukocyte recruitment to sites of inflammation. VCAM is an inducible transmembrane glycoprotein which has a basal level expression in resting endothelial cells but is rapidly expressed upon exposure to pro-inflammatory cytokines (eg IL-1, TNF α). The interaction of VCAM with leukocytes is via the very late antigen 4 (VLA-4) expressed at the leukocyte cell surface. Therefore endothelial cell expression of VCAM functions to induce the infiltration of VLA-4 presenting leukocytes to sites of inflammation which augments rejection responses to allografts or xenografts.

It is believed that porcine VCAM plays an important role in allowing the migration of human leukocytes across porcine endothelial cell monolayers. There is a rationale for believing that blocking this interaction will have beneficial consequences on xenograft survival. Pig VCAM, cloned in 1994, has significant homology with human VCAM(1). As well as the data presented in (1), there is a wealth of evidence from other *in vitro* studies suggesting that pig VCAM interacts efficiently with human leukocyte- expression counter receptor, VLA-4. For instance, in static adhesion assays, antibodies to VCAM significantly inhibit the binding of human NK and T cells to pig endothelium. With NK cells, this disruption inhibits cell lysis which normally results after adhesion to porcine endothelial monolayers.

The effect of anti-VCAM antibodies on T cell mediated xenograft rejection mechanisms is more difficult to predict. In some rodent models of allotransplantation, antibodies

against VCAM have been used to prolong allograft survival. In some instances, long term survival and specific tolerance have been described (2,3), although the precise mechanism of action of these studies was not fully elucidated.

5 3.5 Peptide immunisation strategy

Previous *in vivo* studies using synthetic peptides conjugated to carrier molecules as immunogens have demonstrated the ability to generate the production of biologically active antibodies (68). There is now an extensive literature detailing peptide immunisation strategies which demonstrate enhancement of antibody production by carrier presentation(68-72). Thus, appropriate T cell epitopes can be used to prime T cells for subsequent help to B cells. Recent data has been published reporting the production of IgG by self-reactive B cells following immunisation with a self reacting antigen covalently coupled to a carrier molecule (70). Thereby demonstrating that B cell tolerance to self protein can be overcome.

15 As mentioned above, in order to be recognised by T cells, antigen (self or foreign) must be processed and presented by APC. B cells can act as highly potent APC following endocytosis of antigen via IgG receptors . In the presence of a full complement of activation signals (TCR engagement plus costimulation) T cell activation will occur resulting in the subsequent generation of antibody.

20 Peptides from self proteins are processed and presented to T cells in the same manner as foreign proteins, but because of T cell tolerance, presentation of self peptides does not normally result in T cell activation (70). The absence of T cell recognition may therefore explain, in part, why potentially reactive B cells fail to respond.

The ability to overcome B cell non-responsiveness to self peptides has recently been demonstrated by Dalum *et al* (69). An autoantibody response was generated by the provision of additional T cell help in the form of a strong foreign carrier T cell epitope.

30 Further studies have demonstrated that synthetic peptides conjugated to T cell carrier

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molecules are capable of overcoming B cell non-responsiveness if significant numbers of self-reactive B cells are present in the host (69,70). Insertion of a single foreign T cell epitope into the sequence of Ubiquitin, elicited strong autoantibody production directed against the native molecule (69). In an elegant study by Sad, using GnRH as a self protein
5 chemically linked to diphtheria toxoid (DT) as the synthetic T cell epitope, autoantibodies were produced with specificity for native GnRH (71,72). Following the initial vaccination, the continued presence of the native GnRH *in vivo* maintained the production of Ab. Continued antibody production caused sterility in the immunised mice due to the sustained anti-GnRH antibody response maintained by the continued presence
10 of the native molecule against which the specific B cells were producing antibody. The DT carrier provoked a helper T cell response to assist GnRH specific B cells and break B cell tolerance.

15 4. STATEMENTS OF INVENTION

In its broadest aspect the invention relates to the immunisation of a mammal, preferably a human, with an immunogen which results in the production of antibodies specific to porcine epitopes expressed, typically, but not exclusively, by porcine endothelial cells which are involved in mediating xenograft tissue/organ immune rejection.

20 Immunogen is herein construed as any epitope or combination of epitopes capable of invoking an immune response. The epitope may be T cell specific or B- cell specific. In this context, epitope is construed as any polypeptide, peptide, modified polypeptide, modified peptide (eg typically modification may be by glycosylation or phosphorylation
25 of the epitope).

Typically, the invention encompasses epitopes derived from porcine molecules which are selected from at least one of: CD40; B7.1; B7.2; VCAM.

30 It will be apparent to one skilled in the art that the invention provides means to immunise an individual, ideally prior to xenotransplantation, with an immunogen to a part of a

porcine molecule which contains a B-cell epitope not present in the homologous mammalian polypeptide to ensure the selective production of antibodies to the porcine polypeptide without the development of antibodies to the patients own functional equivalent and without the development of CD4 T cell responses thereby avoiding cell mediated rejection. In addition the immunogen provides blocking antibodies generated by the recipient which abrogate the activity of porcine polypeptides which mediate a rejection response.

It will be still further apparent to one skilled in the art that the invention has significant advantages over prior art attempts to immunosuppress a recipients immune system to porcine cells/tissues. For example, WO 97119971 discloses the use of B7.2 or VCAM polypeptides to produce diagnostic and therapeutic antibodies to monitor transplantation rejection and to block xenotransplant rejection.

This has significant disadvantages. The treatment of a transplant patient with an antibody to, for example VCAM or B7.2, requires periodic administration throughout the life of the patient to maintain the blocking properties of the antibody. Moreover, the immune system will ultimately raise antibodies to the therapeutic antibodies (anti-idiotypic antibodies)resulting in their removal from the patients circulation.

The present invention does not require periodic administration since it is the patients own immune system that is responsible for the production of blocking antibodies to porcine polypeptides. The immune system will not recognise these antibodies as foreign and will therefore not result in the production of anti-idiotypic antibodies.

The present invention involves the use of a foreign T cell epitope to exert significant influences on subsequent responses to molecules conjugated to the carrier. By such means autoantibody responses may be directed against porcine polypeptides in a xenotransplantation context.

According to the present invention there is provided a method of improving the tolerance of an animal, including a human being, to a xenograft, the animal having T cell mediated immunity, the method comprising causing the animal to raise an antibody against a xeno-
molecule involved in the generation of a rejection response in the animal, said antibody
5 being raised by immunising the animal with a chimeric peptide comprising a T cell epitope against which the animal has immunity and a B cell epitope of said xenomolecule.

Accordingly, xenograft specific tolerance is induced in transplant recipients by targeting
10 the direct T cell mediated response by the use of chimeric peptide constructs to stimulate the generation of specific anti-graft tolerance-promoting antibodies by the recipient prior to transplantation. By way of example, the chimeric peptides comprise a T cell epitope conjugated to sequences of porcine polypeptides, B7-1, B7-2, CD40, VCAM. The presence of the engrafted tissue will then serve to maintain and perpetuate the production
15 of antibody by the recipient's B cells.

The present invention also provide a chimeric peptide comprising a T cell epitope and a B cell epitope, said T cell being that of an animal, including a human being of a first species and said B cell being of an animal of a second species, said first and second species such
20 that xeno transplantations suitable from an animal of said second species to an animal of said first species.

In addition, the present invention provides the use of a chimeric peptide improving the tolerance of an animal, including a human being, to a xenograft, the chimeric peptide
25 being as defined above.

According to a further aspect of the invention said immunogenic composition comprises at least one T- cell epitope and at least one B- cell epitope characterised in that said B – cell epitope is derived from at least one porcine polypeptide involved in mediating

xenograft rejection and said T cell epitope is derived from a molecule to which the recipient is already immune.

In yet a further preferred embodiment of the invention said immunogenic composition
5 comprises at least one peptide antigen derived from at least one of porcine: CD40; VCAM; CD86; CD80.

Preferably said peptide antigen is derived from porcine CD40. Ideally said peptide is derived from the amino- terminal domain of porcine CD40, or at least that part of the
10 amino terminal domain that is exposed at the cell surface of a porcine cell presenting CD40. More ideally still said peptide antigen is selected from the peptide sequences presented in Figure 22

Preferably said peptide antigen is derived from porcine VCAM. Ideally said peptide is
15 derived from the amino- terminal domain of porcine VCAM, or at least that part of the amino terminal domain that is exposed at the cell surface of a porcine cell presenting VCAM. More ideally still said peptide antigen is selected from the peptide sequences presented in Figure 24

Preferably said peptide antigen is derived from porcine CD86. Ideally said peptide is
20 derived from the amino- terminal domain of porcine CD86, or at least that part of the amino terminal domain that is exposed at the cell surface of a porcine cell presenting CD86. More ideally still said peptide antigen is selected from the peptide sequences presented in Figure 26.

25 Preferably, said peptide antigen comprises at least 9 amino acid residues. More ideally still said peptide comprises 10 – 30 amino acid residues.

According to a further aspect of the invention there is provided an immunogenic
30 composition according to any previous aspect or embodiment of the invention wherein

said composition further comprises at least one agent capable of enhancing the immune response to said immunogenic composition.

In a preferred embodiment of the invention said agent is a carrier / adjuvant.

5

It is well known in the art that carriers/adjuvants are useful in promoting immune responses to selected antigens. These adjuvants are either crosslinked or coupled to the antigen or co-administered to the animal with the antigen. Adjuvants useful in promoting immune responses are detailed in Vaccine Design: The Subunit and Adjuvant Approach Chapter 7, p141- 228, Plenum Press, New York, 1995. Various carriers, excipients or diluants are available in which said immunogenic composition can be stored and/or administered. For example, and not by way of limitation, the encapsulation of the immunogenic composition in liposomes is a conventional practice. Liposomes are phospholipid based vesicles which are useful as carrying agents for immunogenic compositions and the like.

10
15

According to yet a further aspect of the invention there is provided an antibody, or at least the effective part thereof, directed to at least one region of at least one porcine polypeptide according to the invention.

20

In a preferred embodiment of the invention said antibody is a monoclonal antibody, or at least the effective part thereof. Ideally said antibody is labelled.

It will be apparent to one skilled in the art that antibodies according to the invention will have utility with respect to monitoring the expression of porcine polypeptides presented by porcine tissues/organs.

25

According to a further aspect of the invention there is provided a method to monitor the immune status of a mammalian recipient of a xenograft. Preferably said monitoring method is *in vitro*.

30

According to yet a further aspect of the invention there is provided a method to improve the tolerance of an animal to a xenograft comprising:

- 5 i) administering at least one immunogenic composition according to any previous aspect or embodiment of the invention to an animal; optionally
- ii) monitoring the immune status of said animal to said immunogenic composition;
- iii) transplantation of at least one porcine tissue/organ into said animal; and, optionally
- 10 iv) monitoring the animal for a rejection response to said porcine tissue/organ.

In a preferred method of the invention said animal is human.

- 15 In a further preferred method of the invention said xenograft is any vascularised graft and/or immunogenic porcine cell/tissue.

In a further preferred method of the invention said xenograft is porcine pancreatic islets.

- 20 It will be apparent to one skilled in the art that (ii) above can be conducted either by monitoring for the presence of antibodies to co-stimulatory molecules in sera (for example by ELISA or by FACS analysis of cells expressing said co-stimulatory molecules), or alternatively, or in addition, monitoring the presence of cytolytic T- cells in the blood of the treated animal by conventional T- cells lysis assays.

- 25 The potential benefits of the use of a chimeric peptide of the invention are that it avoids the need for injection of blocking antibodies or fusion proteins. Furthermore, the induction of a recipient antibody response circumvents the problems most commonly associated with administration of xenogeneic antibodies or fusions proteins, namely the immune response against the administered reagent.

30

An embodiment of the invention will now be described, by example only and with reference to the following Tables and Figures;

5 Table 1 represents the regions of non-homology in human CD40 with respect to the homologous porcine CD40;

Table 2 represents the regions of non-homology in human VCAM with respect to the homologous porcine VCAM;

10 Table 3 represents the regions of non-homology in human CD86 with respect to the homologous porcine CD86;

Figure 1a is a diagrammatic representation of direct xenorecognition and Figure 1b is a diagrammatic representation of indirect xenorecognition;

15 Figure 2 represents the porcine CD86 nucleic acid sequence;

Figure 3 represents the porcine CD86 cDNA sequence obtained by reverse transcription of porcine mRNA followed by PCR amplification;

20 Figure 4 represents a comparison of the nucleotide sequence of the cDNA in Figure 2 with the published porcine CD86 sequence;

Figure 5 represents a comparison of the cDNA sequence in Figure 2 with the published murine and human CD86 sequences;

25

Figure 6 represents the translated amino acid sequence of the cDNA in Figure 2 compared with porcine, human and murine amino acid sequences;

30 Figure 7 represents the position of porcine B7.1 oligonucleotide primers with respect to the human and murine B7.1 nucleic acid sequences;

Figure 8a represents a comparison of the human, murine and bovine CD40 nucleic acid sequences; Figure 8b represents a comparison of the human, murine and bovine CD40 amino acid sequences;

5

Figure 9 represents FACS analysis of the expression of CD86 (B7.2) after transfection with a vector encoding porcine CD86 (B7.2);

Figure 10 represents FACS analysis of the expression of CD86 (B7.2) by transiently transfected cells with a vector encoding porcine CD86(B7.2);

10

Figure 11 represents flow cytometric analysis of cells transfected with porcine CD86(B7.2);

Figure 12 represents the position of nine CD86(B7.2) derived peptides in the porcine CD86(B7.2) sequence;

15

Figure 13 represents a comparison of T cell proliferation response to whole ovalbumen or the ovalbumen peptide Ova₃₂₃₋₃₃₉;

20

Figure 14a represents the differential binding of B7.2 specific peptide sera or ovalbumen control sera by peptide ELISA;

Figure 14b represents the in vitro recognition of B7.2 derived peptides 4 and 6 by mouse sera immunised with peptides 4 or 6;

25

Figure 15a represents the in vitro recognition of the B7.2 peptide sera and control ova peptide sera by peptide ELISA;

Figure 15b represents the inhibition of direct mouse anti porcine T cell responses by peptide 4 and 6 sera which also shows no inhibition of of costimulation by murine CD86;

Figure 16 represents the differential binding of the B7.2 derived peptide 4 sera or ova control peptide sera by peptide ELISA;

Figure 17a represents flow cytometric analysis of P815 cells transfected with porcine CD86 following staining with sera from peptide 4 or control ova peptide sera;

Figure 17b represents FACS analysis of P815 cells transfected with porcine CD86 or CHO cells transfected with murine CD86 following staining with sera from mice sera derived from peptide 4 or peptide 6;

Figure 18 represents a preparation of porcine pancreatic islets isolated from a large white pig;

Figure 19 is a schematic representation of the chimeric peptide immunisation and transplantation protocol;

Figure 20 shows that anti-porcine CD86 antisera prolongs the survival of transplanted porcine pancreatic islets;

Figure 21 is a comparison of the amino acid sequence of porcine and human CD40 (underlined sequences are peptides identified in table 1);

Figure 22 is the translated amino acid sequence of porcine CD40 (underlined sequences are peptides identified in table 1);

Figure 23 is a comparison of the amino acid sequence of porcine and human VCAM (underlined sequences are peptides identified in table 2);

Figure 24 is the translated amino acid sequence of porcine VCAM (underlined sequences are peptides identified in table 2);

Figure 25 is a comparison of the amino acid sequence of porcine and human CD86
5 (underlined sequences are peptides identified in table 3); and

Figure 26 is the translated amino acid sequence of human CD86 (underlined sequences are peptides identified in table 3)

10 5. SPECIFIC EMBODIMENTS

5.1 Cloning porcine costimulatory molecules

5.1.1 Cloning porcine B7-2

RNA was extracted from primary and transformed porcine cells using a standard protocol. mRNA was then reverse transcribed and porcine B7-2 (poB7-2) amplified from
15 the cDNA by 35 cycles of PCR at 56⁰ C with 1.5mM magnesium. The 5' and 3' primers GCATGGATCCATGGGACTGAGTAACATTCTCTTTG and GCATGTCGACTTAAAAATCTGTAGTACTGTTGTC respectively were designed on the basis of the published poB7-2 sequence (60) to overlay the start and stop codons (Figure 2). A 956 base pair fragment was generated and subcloned into the BamH1 & Sall restriction sites of pbluescript. The nucleotide sequence was determined using
20 standard m13 forward and reverse primers. The sequence of a single clone, CD86(i) is illustrated in Figure 3, with comparison to the published sequences from porcine (Figure 4), human and murine B7-2 (Figure 5). One base pair difference is detected between our clone, CD86(i), and the published sequence at the 3' prime end. This, however, is
25 unlikely to be an important difference with respect to either poB7-2 expression or binding to its ligand. The predicted amino acid sequence of CD86(i) , compared to that of porcine, human and mouse B7-2 is shown in Figure 6.

5.1.2 Cloning porcine B7-1 and CD40

RNA extracted from phytohaemagglutinin (PHA) or poke-weed mitogen (PMW) stimulated porcine PBMC and transformed porcine endothelial cells is being used to amplify cDNA encoding the costimulatory molecules B7-1 and CD40. B7-1 Primers were designed on the basis of conserved areas following comparison of murine and human (29,49) sequences. External (lying outside the coding region) AGACCGTCTTCCTTTAG(3'i), TTGGATCCTCCATGTTATCCC (3'ii) and AGCATCTGAAGC (5') and internal (within the coding region) ATGGATCCTCCATTTTCCAACC (3') and TTGTCGACATCTACTGGC (5') primers have been designed as depicted in Figure 7. The generation of two 3' primers is due to significant differences between the human and murine sequences in the terminal coding regions. Resulting PCR fragments will be subcloned as described above using the restriction sites BamHI and SalI contained within the promoter sequence. Constructs will then be sent for sequence confirmation.

CD40 primers were designed in a similar manner following sequence alignment of published CD40 sequences from human, mice and cattle (73,74,75) as illustrated in Figures 8A & B. The 5' and 3' primer sequences are GGATCCTCACTGTCTCTCCTGCACTGAGATGCGACTCTCCTCTTTGCCGTCCG TCCTCC and GAATTCATGGTTCTGTTGCCTCTGCAGTG respectively containing the BamHI and EcoRI restriction sites.

5.2 Generation of porcine costimulatory molecule expressing cell transfectants

The poB7-2 molecule (CD869(i)) has been subcloned into the eukaryotic expression vector pci.neo carrying the neomycin drug-selectable marker. This is being used to transfect M1 and M1.DR1 transformed murine cell lines using a standard calcium phosphate precipitation method. G418 resistant pci.neo expressing cells will be selected using dynabead purification and highly expressing clones is selected by limiting dilution.

Stable poB7-2 M1 and P815 transfectants have been generated by this approach using the poB7-2 DNA construct supplied to us by Maher *et al* (Figure 9). transient transfections of M1 and P815 cells have been generated using our CD86(i) construct (Figure 10).

3 particular assays are undertaken using the CD86(i) transfected cells.

- 5 (I) comparative costimulatory function of poB7-2 with human B7-1 in the context of MHC restriction;
- (II) flow cytometric analysis of specific anti-poB7-2 antibodies in the sera of immunised mice; and
- (III) generation of specific anti-poB7-2 monoclonal antibodies.

10

(I) Comparative *in vitro* analysis is performed to determine the costimulatory function of poB7-2 or poB7-1 in the context of the human MHC class II molecule HLA-DR1, with that of human B7-1 or B7-2 in the context of DR1, in proliferation assays with human or porcine responders.

15

(II) Transfected P815 cells are crucial reagents for the detection of porcine anti-B7-2 antibody in the sera of immunised mice which have undergone the chimeric peptide immunisation regimen. Flow cytometric analysis with control or poB7-2 -transfected P815 cells, reflects the specificity of sera for B7-2. Preliminary studies with C57BL-6 mice immunised with a pool of all nine B7-2 peptides have demonstrated the preferential binding of B7-2 peptide sera to porcine B7-2 transfected P815 cells (Figure 11a and 11b).

20

(III) Mab with specificity for poB7-2 are generated by immunisation of Balb/c mice with poB7-2 expressing P815 cells . The spleens from immunised mice are fused with the NS0 fusion partner and successful fusion's selected by virtue of HAT selection. Flow cytometric staining of poB7-2 P815 transfectants with culture supernatants enable the identification of MAb secreting cells. Cells are grown in culture and the medium harvested for antibody purification by passage over Protein G following ammonium sulphate precipitation. Techniques for the preparation on monoclonal antibodies are well

25

known in the art and with reference to publications such as Harlow and Lane Antibodies; A Laboratory Manual; Cold Spring Harbour Laboratories.

- 5 MAb with specificity for B7-1 and CD40 are generated using the same protocol. These MAb will provide valuable reagents for further characterising the expression of CS molecules on relevant porcine tissues.

5.3 Design and synthesis of poB7-2/OVA chimeric peptide constructs

- 10 Nine different peptides derived from the sequence of poB7-2 were initially selected for synthesis. Porcine B7-2 peptides, 6-22mer in size, were selected as determined by the predicted size of a B cell epitope. Peptides were selected for synthesis in combination with a T cell epitope OVA 323-339. B7-2 peptides were selected on the basis of 3D computer modelling (in collaboration with Paul Travers) and on the basis of predicted antigenicity and hydrophilicity using the SeqAid II computer software package. All of the
- 15 nine peptides reflect linear epitopes. The positions of the nine peptides in the cloned poB7-2 sequence are indicated (Figure 12). Synthetic peptide sequences are detailed in Table 1

Table 1

Peptide Name	Peptide Sequence	Position
Peptide 1	ISQAVHAAHAEINEAGRSFDQATWTLR	81-90
Peptide 2	ISQAVHAAHAEINEAGRLPCHFTNSQ	32-40
Peptide 3	ISQAVHAAHAEINEAGRKGPHGLVPIHQMS	109-121
Peptide 4	ISQAVHAAHAEINEAGRGLVPIHQMS	113-121
Peptide 5	ISQAVHAAHAEINEAGRVQIKDKGSYQC	94-104
Peptide 6	ISQAVHAAHAEINEAGRCSTQGYPEPQR	151-162
Peptide 8	ISQAVHAAHAEINEAGRKSQAYFNETGEL	21-32
Peptide 9	ISQAVHAAHAEINEAGRSLKSQAYFNET	17-29
Peptide 10	ISQAVHAAHAEINEAGRYMGRTSFDQATWT	76-88
Ova Peptide	ISQAVHAAHAEINEAGR	323-339

- 5 The peptide sequences and amino acid positions for peptides 1-10 relate to the position of the B7-2 peptide sequence within porcine B7-2. The amino acid position for the ova sequence is only indicated for the Ova peptide. A 17 amino acid peptide from chicken egg albumin (ovalbumin) was selected as the T cell epitope, OVA323-339 (ISQAVHAAHAEINEAGR). This epitope was selected on the basis of published reports
- 10 for the generation of a H-2^b restricted T cell response (76,77). We have demonstrated the ability of C57BL-6 mice (H-2^b haplotype) to mount a proliferative response to both the native molecule and to the OVA 323-339 peptide following immunisation with whole ovalbumin (Figure 13). Peptides were generated on a peptide synthesiser (Genosys) and crude peptides were purified by HPLC to greater than 70% purity. Sera from OVA
- 15 control immunised mice should ideally not recognise the 323-339 sequence, indicating that the T cell epitope is devoid of B cell determinants.

5.4 Tolerance induction

5.4.1 *In vivo* tolerance induction strategy

- 20 C57BL-6 mice are immunised with whole ovalbumin in CFA, followed by either control peptide (OVA peptide) or CS peptides (OVA-B7-2 constructs) for three weekly immunisations. Blood is collected following sacrifice and sera prepared using a standard

technique. Presence of specific mouse anti-porcine B7-2 IgG and/or IgM Ab is detected by one of two strategies.

Peptide ELISAs are used to screen for the presence of anti-peptide antibody in the sera.

- 5 Peptides are coated to plates by virtue of aldehyde linkages to allow free access of Ab to the peptide (78). Plates are coated with individual peptides or the ova control peptide to enable the identification of specific peptides of interest. To detect reactivity of sera with the native B7-2 molecule expressed on the surface of PoB7-2 transfected P815 cells, flow cytometry is performed following surface staining. Having identified CS peptide of interest (peptide ELISA positive and recognising native B7-2) the sera is used to inhibit *in vitro* T cell proliferative responses. This determines whether the antibody is a blocking antibody.

- 15 *In vivo* studies are performed using the islet transplant system. Antibodies which recognise the native molecule but fail to block a proliferative response are useful polyclonal antibody reagents.

- Immunisations involved two groups of mice, one received a pool of all nine B7-2 peptides, and one receiving ova control peptide. The harvested sera were screened by peptide ELISA (Figure 14a or 14b) which enabled the identification of peptides of interest. Antisera to peptides 2, 4 and 6 clearly demonstrate preferential binding to B7 peptide than to ova control. The sera has also demonstrated enhanced binding to poB7-2 transfected cells (Figure 11). Peptide 4 and 6 were selected as candidate peptides and used in subsequent immunisation protocol. Immunisation with peptide 4 or 6 clearly produced a significant level of IgG with specificity for peptides 4 and 6 in the sera of immunised mice (Figure 15a and 15b). The specificity of the sera for peptide 4 and not to ova control is demonstrated in Figure 16. The ability of sera from peptide 4 and 6 immunised mice to specifically recognise the native porcine B7-2 molecule expressed on the surface of porcine B7-2 transfected P815 cells is illustrated in Figure 17a and 17b.
- 30 Untransfected control P815 cells do not stain with the Peptide 4 or 6 sera, neither do

control or transfected cells incubated with ova peptide sera. Similar protocols will be followed with peptide 2. These data clearly demonstrate the ability of this technique to generate anti-peptide antibody directed against an amino acid sequence, by virtue of a carrier T cell epitope.

5

An identical strategy will be followed with peptides designed on the basis of porcine CD40 and porcine B7-1 once the DNA sequence encoding these molecules has been elucidated.

10 5.4.2 Functional assessment; prolongation of pancreatic islet xenograft survival

Islet xenografts being non-vascular are rejected solely by T cell mediated mechanisms (79,80), thereby providing an ideal system to study modulation of T cell mediated reactions, please see Figure 18. A very clear role for cell mediated rejection of islets has been demonstrated and is reported to be greater than the comparable alloresponse (80).

15 Transplantation of porcine pancreatic islets to mice is an established procedure, which is well documented in the literature (80-83). Studies within this laboratory have demonstrated a decrease in hyperglycaemia (Figure 18) following transplantation of pancreatic islets from large white pigs under the kidney capsule of C57BL-6 mice rendered diabetic by intraperitoneal administration of streptozotocin, please see Figure 19
20 and 20. Further optimisation of the isolation procedure (84,85) is required to enable purification of fully functional islets. Transplanted islets usually survive between 6-10 days in the absence of any immunosuppression. Successful modulation of direct T cell mediated xenorejection will be monitored by prolongation of islet survival beyond day 10, with comparison to the appropriate controls.

25

The results obtained with B7-2 to date, demonstrate the ability of synthetic B7-2 peptides conjugated to a known T cell helper epitope to generate the production of anti-porcine B7-2 antibody *in vivo*. These antibodies if directed towards the binding site between B7
30 isoforms and CD28, in association with antibodies directed against CD40-CD40L will

block the costimulation of human T cells with direct anti-pig xenoreactivity thereby prolonging islet survival in a xenotransplantation context.

Having established the suitability of such an approach in a pig islet to mouse *in vivo* model, studies would progress to pig to primate transplantation systems prior to clinical trials.

5.5 Adaptations for clinical use of these strategies

For clinical applicability the following requirements are necessary:

- 10 (I) selection of a suitable T cell epitope to replace OVA. One candidate molecule is tetanus toxoid (TT) which is a widely used antigen for use in human immunisation strategies (68,86). The prior immunisations of most adults with TT is an additional benefit to this strategy as memory T cells are already present in the circulation.
- (ii) An efficient and rapid screening method is used to detect the presence of anti-donor
- 15 (pig) B7-2 antibodies in the absence of a specific B7-2 directed T cell response generated by the recipient which would accelerate graft rejection.

6. SUMMARY OF SPECIFIC EMBODIMENTS

- 20 The above examples relate to a novel strategy to inhibit costimulation by porcine cells of human T cells with direct anti-pig xenoreactivity. This is of particular importance in the context of xenotransplantation of porcine organs due to the expression of costimulatory molecules on porcine endothelial, as well as bone marrow-derived antigen presenting cells.

25

- Recipients are immunised with hybrid synthetic peptides comprising a T cell epitope conjugated to sequences of the porcine costimulatory molecules, CD80, CD86 and CD40. Peptides that induce antibodies specific for regions of the costimulatory molecules involved in binding to their counter-receptors on human cells (CD28 and CD154) are
- 30 therefore capable of blocking the delivery of costimulation. Once the antibody response has been induced, the transplanted organ will recall this response due to the expression of

the costimulatory molecules, thereby sustaining this response, and providing an endogenous mechanism of costimulatory blockade.

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CD86 (B7-2)

Human and porcine CD86 protein sequences were aligned and regions of non-homology identified. We predict that the peptide sequences will be derived from those regions listed below or from any overlap regions between any of these peptides.

The sequences of predicted interest for containing potential antibody epitopes have been selected on the basis of less than 75% sequence identity.

Region	Position	% sequence identity
i	18-42	72%
ii	55-73	55%
iii	101-127	63%
iv	136-165	56%

Regions (iii) and (iv) encompass those containing the peptide 4 and 6 sequences identified in mice.

CD40

Human and porcine CD40 protein sequences were aligned and regions of non-homology identified. We predict that the peptide sequences will be derived from those regions listed below or from any overlap regions between any of these peptides.

The sequences of predicted interest for containing potential antibody epitopes have been selected on the basis of less than 75% sequence identity.

Region	Position	% sequence identity
i	25-48	63%
ii	49-75	74%
iii	93-114	59%
iv	123-139	63%
v	158-176	68%
vi	208-227	45%
vii	231-248	21%

VCAM-1

Human and porcine VCAM-1 protein sequences were aligned and regions of non-homology identified. We predict that the peptide sequences will be derived from those regions listed below or from any overlap regions between any of these peptides. The sequences of predicted interest for containing potential antibody epitopes have been selected on the basis of less than 75% sequence identity.

Region	Position	% sequence identity
i	1-15	44%
ii	16-33	63%
iii	49-65	58%
iv	74-85	42%
v	100-117	50%
vi	122-140	56%
vii	144-157	64%
viii	162-191	47%
ix	209-221	62%
x	290-301	67%
xi	322-342	62%
xii	362-379	67%
xiii	448-465	67%

CLAIMS

1. A method of improving tolerance to a porcine xenograft comprising immunising a mammal with an immunogen comprising:
 - i) a T- cell epitope; and
 - ii) a B-cell epitope characterised in that the B-cell epitope is a porcine polypeptide involved in mediating xenograft rejection and derived from a region of a porcine polypeptide which has less than 75% sequence identity to the corresponding region of the equivalent human polypeptide.
2. A method according to Claim 1 wherein the B-cell epitope is a peptide derived from at least one porcine polypeptide selected from; CD40; CD80; CD86 or VCAM.
3. A method according to Claim 1 or 2 wherein the peptide is selected from at least one peptide represented in Figure 22.
4. A method according to Claim 1 or 2 wherein the peptide is selected from at least one peptide represented in Figure 24.
5. A method according to Claim 1 or 2 wherein the peptide is selected from at least one peptide represented in Figure 26.
6. A method according to any of Claims 1-5 wherein the T – cell epitope is derived from tetanus toxoid polypeptide.
7. A composition comprising an immunogen characterised in that the immunogen has a T – cell epitope and a B- cell epitope wherein the B – cell epitope is derived from a region of a porcine polypeptide which has less than 75% sequence identity to the corresponding region of the equivalent human polypeptide.

8. A composition according to Claim 7 wherein the porcine polypeptide is expressed by vascular endothelial cells of said xenograft.
9. A composition according to Claims 7 or 8 wherein the B-cell epitope is derived from at least one porcine polypeptide selected from; CD40; CD86; CD80; VCAM.
10. A composition according to Claim 9 wherein the B- cell epitope is selected from at least one peptide as represented in Figure 22 .
11. A composition according to Claim 9 wherein the B- cell epitope is selected from at least one peptide as represented in Figure 24 .
12. A composition according to Claim 9 wherein the B- cell epitope is selected from at least one peptide as represented in Figure 26.
13. A composition according to Claims 9 or 12 wherein the B- cell epitope is derived from the extracellular domain of CD86.
14. A composition according to any of Claims 7 - 13 wherein the T- cell epitope is derived from tetanus toxoid.
15. A composition according to any of Claims 7 - 14 wherein the composition further comprises a carrier capable of enhancing the immune response to said immunogen.
16. An antibody, or the effective part thereof, characterised in that said antibody is capable of binding to a region of a porcine polypeptide which has less than 75% sequence identity to the corresponding region of the equivalent human polypeptide.
17. An antibody according to Claim 16 wherein the antibody is a monoclonal antibody.

18. An antibody according to Claims 16 or 17 wherein the antibody is modified with at least one detectable label.
19. A method to monitor the immune status of a mammalian recipient of a xenograft comprising:
- i) removing a sample from a xenograft recipient to be tested;
 - ii) contacting said sample with the antibody according to Claims 16 -18; and
 - iii) monitoring the expression of a porcine polypeptide involved in mediating xenograft rejection.
20. A method to treat a mammal prior to receiving a xenograft comprising:
- i) immunising a mammal with a composition according to Claims 7-15;
 - ii) assessing the immune status of said mammal to said immunogenic composition;
 - iii) transplantation of said xenograft tissue/organ into a recipient mammal; and
 - iv) monitoring the rejection response to said xenograft.
21. A method according to Claim 19 or 20 wherein the xenograft is of porcine origin and said mammal is human.
22. A method according to any of Claims 19 -21 wherein the xenograft is at least one vascularised graft and/or immunogenic porcine cell/tissue.
23. A method according to any of Claims 19 – 22 wherein the xenograft is pancreatic islets.

PCT

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(54) Title: IMPROVEMENT OF TOLERANCE TO A XENOGRRAFT			
(57) Abstract <p>The invention hereindescribed relates to a method to improve the tolerance of a mammal, preferably a human, to a xenograft through immunisation of the recipient mammal with an immunogen comprising both a B cell epitope derived from porcine polypeptides and T cell epitope. The invention also encompasses immunogenic compositions comprising said immunogens and methods to monitor the status of the xenograft.</p>			

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Figure 1

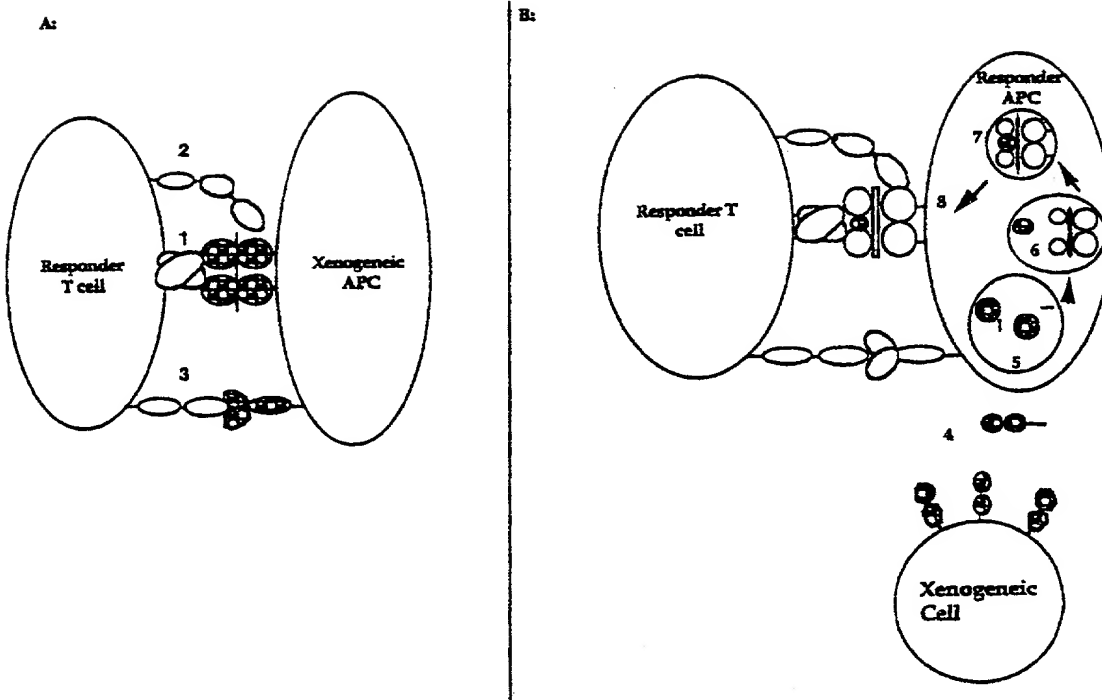


FIGURE 2

GCATGGATCCATGGGACTGAGTAACATTCTCTTTG

1 **ATGGGACTGAGTAACATTCTCTTTGTGATGGTCCTCCT**

39 GCTCTCTGGTGCTGCCTCCTTGA~~AA~~AAGTCAGGCATATTTCAATGAGA

86 CTGGAGAACTGCCGTGCCATTTTACA~~AA~~ACTCGCAGAACCTAAGCCTG

133 GATGAGCTGGTCATATTTTGGCAGGACCAGGATAACCTGGTTCTCTA

181 CGAGCTATACCGAGGCCAAGAGAAGCCTCATAATGTTAATTCCAAG

227 TATATGGGTCGCACAAGCTTTGACCAGGCCACCTGGACCCTGAGACT

274 CCACAACGTTCAAATCAAGGACAAGGGCTCATATCAATGTTTCATC

321 CATCATAAAGGGCCGCATGGACTTGTTCTATCCACCAGATGAGTTC

368 TGACCTATCATTGCTTGCTAACTTCAGTCAACCTGAAATAAACCTAC

415 TTACTAATCACACAGAAAATTCTGTCATAAATTTGACCTGCTCATCT

462 ACACAAGGCTACCCAGAACCCCAGAGGATGTATATGTTGCTAAATA

509 CGAAGAATTCAACCACTGAGCATGATGCTGACATGAAGAAATCTCA

556 AAATAACATCACGGA~~ACTCTACAATGIATCAATCAGGGTGTCTCTT~~

602 CCCATCCCTCCCGAGACAAATGTGAGCATCGTCTGTGTCCTGCAACTT

649 GAGCCAAGCAAGACACTGCTTTTCTCCCTACCTTGTAATATAGATGC

696 AAAGCCACCTGTGCAACCCCCCTGTCCCAGACCACATCCTCTGGATTGC

743 AGCTCTACTTGTAACAGTGGTCGTTGTGTGTGGGATGGTGTCTTTGT

790 AACACTAAGGAAAAGGAAGAAGAAGCAGCCTGGCCCCCTCTAATGA

837 ATGTGGTGAAACCATCAAAATGAACAGGAAGGCGAGTGAACAAAC

884 TAAGAACAGAGCAGAAGTCCATGAACGATCTGATGATGCCCAGTGT

931 GATGTTAATATTTTAAAGACAGCCTCAGATGACAACAGTACTACAG

GACAACAGTACTACAG

978 **ATTTTTTAATTAAAGAGTAAACTCC**

ATTTTTTAAGTCGACATGC

Figure 3

1 CACCGCGGTG CGGCCGCTCT AGAACTAGTG GATCCATGGG ACTGAGTAAC
51 ATTCTCTTTG GGATGGTCCT CCTGCTCTCT GGTGCTGCCT CCTTGAAAAG
101 TCAGGCATAT TTCAATGAGA CTGGAGAACT GCCGTGCCAT TTTACAAACT
151 CGCAGAACCT AAGCCTGGAT GAGCTGGTCA TATTTTGGCA GGACCAGGAT
201 AACCTGGTTC TCTACGAGCT ATACCGAGGC CAAGAGAAGC CTCATAATGT
251 TAATTCCAAG TATATGGGTC GCACAAGCTT TGACCAGGCC ACCTGGACCC
301 TGAGACTCCA CAACGTTCAA ATCAAGGACA AGGGCTCATA TCAATGTTTC
351 ATCCATCATA AAGGGCCGCA TGGACTTGTT CCTATCCACC AGATGAGTTC
401 TGACCTATCA GTGCTTGCTA ACTTCAGTCA ACCTGAAATA AACCTACTTA
451 CTAATCACAC AGAAAATTCT GTCATAAATT TGACCTGCTC ATCTACACAA
501 GGCTACCCAG AACCCAGAG GATGTATATG TTGCTAAATA CGAAGAATTC
551 AACCACTGAG CATGATGCTG ACATGAAGAA ATCTCAAAAT AACATCACGG
601 AACTCTACAA TGTATCAATC AGGGTGTCTC TTCCCATCCC TCCCGAGACA
651 AATGTGAGCA TCGTCTGTGT CCTGCAACTT GAGCCAAGCA AGACACTGCT
701 TTTCTCCCTA CCTTGTAATA TAGATGCAAA GCCACCTGTG CAACCCCTTG
751 TCCCAGACCA CATCCTCTGG ATTGCAGCTC TACTTGTAAC AGTGGTCGTT
801 GTGTGTGGGA TGGTGTCTT TGTAACACTA AGGAAAAGGA AGAAGAAGCA
851 GCCTGGCCCC TCTAATGAAT GTGGTGAAAC CATCAAAATG AACAGGAAGG
901 CGAGTGAACA AACTAAGAAC AGAGCAGAAG TCCATGAACG ATCTGATGAT
951 GCCCAGTGTG ATGTTAATAT TTAAAGACA GCCTCAGATG ACAACAGTAC
1001 TACAGATTTT TAAGTCGACC TCGAGGGGGG GCCCGGTACC AGCTTTTGT

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Figure 4: Comparison of the nucleotide sequence of CD86(i) with the published sequence for porcine CD86.

10 20 30 40
ATGGGACTGAGTAACATTCTCTTTGATGGTCTCTCTCTCTGG
.....
CACCGCGGTGCGGCGGCTCTAGAACTAGTGGATCCATGGGACTGAGTAACATTCTCTTTGGGATGGTCTCTCTCTCTGG
10 20 30 40 50 60 70 80

50 60 70 80 90 100 110 120
TGCTGCTCTCTTGAAAAGTCAGGCATATTTCATGAGACTGGGAACTGCCGTGCCATTTTACAACTGGCAGAACCTAAGC
.....
TGCTGCTCTCTTGAAAAGTCAGGCATATTTCATGAGACTGGGAACTGCCGTGCCATTTTACAACTGGCAGAACCTAAGC
90 100 110 120 130 140 150 160

0 140 150 160 170 180 190 200 210
CTGGATGAGCTGGTCATATTTTGGCAGGACCAGGATAACCTGGTCTCTTACGAGCTATACCGAGGCCAAGAGAAGCCTCATA
.....
CTGGATGAGCTGGTCATATTTTGGCAGGACCAGGATAACCTGGTCTCTTACGAGCTATACCGAGGCCAAGAGAAGCCTCATA
170 180 190 200 210 220 230 240

220 230 240 250 260 270 280 290
TGTTAATTCGAAGTATATGGGTGCGACAAGCTTTGACCAGGCCACCTGGACCCCTGAGACTCCACAACGTTCAAATCAAGGA
.....
TGTTAATTCGAAGTATATGGGTGCGACAAGCTTTGACCAGGCCACCTGGACCCCTGAGACTCCACAACGTTCAAATCAAGGA
250 260 270 280 290 300 310 320

300 310 320 330 340 350 360 370
TAGGGCTCATATCAATGTTTCATCCATCATAAAGGGGCCCATGGACTTGTTCCTATCCACCAGATGAGTTCGACCTATCA
.....
TAGGGCTCATATCAATGTTTCATCCATCATAAAGGGGCCCATGGACTTGTTCCTATCCACCAGATGAGTTCGACCTATCA
330 340 350 360 370 380 390 400 410

380 390 400 410 420 430 440 450
GCTTGCTAACTTCAGTCAACCTGAAATAAACCTTCTTACTAATCACACAGAAAATTCGTGTCATAAATTGACCTGCTCAT
.....
GCTTGCTAACTTCAGTCAACCTGAAATAAACCTTCTTACTAATCACACAGAAAATTCGTGTCATAAATTGACCTGCTCAT
420 430 440 450 460 470 480 490

460 470 480 490 500 510 520 530
ACACAAGGCTACCCGGAACCCGAGGAGTGTATATGTTGCTAAATACGAAGAATTCAACCACTGAGCATGATGCTGACAT
.....
ACACAAGGCTACCCGGAACCCGAGGAGTGTATATGTTGCTAAATACGAAGAATTCAACCACTGAGCATGATGCTGACAT

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540 550 560 570 580 590 600 610 620
GAAGAAATCTCAAATAACATCACGGAACCTCTACAATGTATCAATCAGGGTGTCTCTTCCCATCCCTCCCGAGACAAATGTG
.....
GAAGAAATCTCAAATAACATCACGGAACCTCTACAATGTATCAATCAGGGTGTCTCTTCCCATCCCTCCCGAGACAAATGTG
580 590 600 610 620 630 640 650

630 640 650 660 670 680 690 700
AGCATCGTCTGTGTCTCTGCAACTTGAGCCAAGCAAGACACTGCTTTTCTCCCTACCTTGTAATATAGATGCAAAGCCACCTG
.....
AGCATCGTCTGTGTCTCTGCAACTTGAGCCAAGCAAGACACTGCTTTTCTCCCTACCTTGTAATATAGATGCAAAGCCACCTG
660 670 680 690 700 710 720 730

710 720 730 740 750 760 770 780
TGCAACCCCTGTGCCAGACCACATCCTCTGGATTGCAGCTCTACTTGTAAACAGTGGTCTGTGTGTGGGATGGTGTCTCTT
.....
TGCAACCCCTGTGCCAGACCACATCCTCTGGATTGCAGCTCTACTTGTAAACAGTGGTCTGTGTGTGGGATGGTGTCTCTT
740 750 760 770 780 790 800 810 820

790 800 810 820 830 840 850 860
TGTAACACTAAGGAAAAGGAAGAAGAAGCAGCCTGGCCCCCTCTAATGAATGTGGTGAACCATCAAAATGAACAGGAAGGCG
.....
TGTAACACTAAGGAAAAGGAAGAAGAAGCAGCCTGGCCCCCTCTAATGAATGTGGTGAACCATCAAAATGAACAGGAAGGCG
830 840 850 860 870 880 890 900

870 880 890 900 910 920 930 940
GTGAACAACTAAGAACAGAGCAGAAGTCCATGAACGATCTGATGATGCCCACTGTGATGTTAATATTTTAAAGACAGCCT
.....
GTGAACAACTAAGAACAGAGCAGAAGTCCATGAACGATCTGATGATGCCCACTGTGATGTTAATATTTTAAAGACAGCCT
910 920 930 940 950 960 970 980

50 960 970 980 990
AGATGACAACTAGTACTACAGATTTTAAATTAAGAGTAAACTCC
.....
AGATGACAACTAGTACTACAGATTTTAAAGTCACTCCAGGGGGGGCCCGTACCAGCTTTTGT
990 1000 1010 1020 1030 1040 1050

09868605.091201

FIGURE 5

Contig	ACCATGGGACTGAGTAACATTCTCTTTGTGATGGTCTTCTGCTCTCT
Murine B7-2	-CCATGGGACTGAGTAACATTCTCTTTGGGATGGTCTTCTGCTCTCT
Porcine CD68(i)	ACCATGGGCTTGGCAATCCTTATCTTTGTGACAGTCTTGTGATCTCA
Human B7.2	ACTATGGGACTGAGTAACATTCTCTTTGTGATGGCTTCTGCTCTCT

GGTCTGCTTCCBTGAAGABTCAAGCTTATTTCAATGAGACTGCAGAHCTGCCGTGCCAATTTA
GGTCTGCTTCCBTGAAGABTCAAGCTTATTTCAATGAGACTGCAGAHCTGCCGTGCCAATTTA
GATGCTGTTTCCGTGGAGACGCAAGCTTATTTCAATGGGACTGCATATCTGCCGTGCCAATTTA
GGTCTGCTTCCBTGAAGABTCAAGCTTATTTCAATGAGACTGCAGAHCTGCCGTGCCAATTTA

CAAACCTCTCAAAACCTAAGCCTGAGTGAGCTGGTAGTATTTTGGCAGGACCAGGAAAACCTTGGT
CAAACCTCTCAAAACCTAAGCCTGAGTGAGCTGGTAGTATTTTGGCAGGACCAGGATAACCTGGT
CAAAGGCTCAAAACCTAAGCCTGAGTGAGCTGGTAGTATTTTGGCAGGACCAGGAAAACCTTGGT
CAAACCTCTCAAAACCTAAGCCTGAGTGAGCTGGTAGTATTTTGGCAGGACCAGGAAAACCTTGGT

TCTGTACGAGCTATACCTTAGGCAAAGAGAACTTGATAGTGTAAATCCAGTATATGGGCCGC
TCTGTACGAGCTATACCTTAGGCAAAGAGAACTTGATAGTGTAAATCCAGTATATGGGCCGC
TCTGTACGAGCTATACCTTAGGCAAAGAGAACTTGATAGTGTAAATCCAGTATATGGGCCGC
TCTGTACGAGCTATACCTTAGGCAAAGAGAACTTGATAGTGTAAATCCAGTATATGGGCCGC

ACAAGCTTTGACHVGGACAVCTGGACCTGAGACTTCACAATGTTTCAGATCAAGGACAAGGGCT
ACAAGCTTTGACHVGGACAVCTGGACCTGAGACTTCACAATGTTTCAGATCAAGGACAAGGGCT
ACGAGCTTTGACAGGAACAACCTGGACTCTACGACTTCACAATGTTTCAGATCAAGGACAAGGGCT
ACAAGCTTTGACHVGGACAVCTGGACCTGAGACTTCACAATGTTTCAGATCAAGGACAAGGGCT

CGTATCAATGTTTTCATCCATCAHAAAVVGGCCACAGGAHTDATTCBCATCCACCAGATGADTTC
CATATCAATGTTTTCATCCATCAHAAAVVGGCCACAGGAHTDATTCBCATCCACCAGATGADTTC
CGTATCAATGTTTTCATCCATCAHAAAVVGGCCACAGGAHTDATTCBCATCCACCAGATGADTTC
TGTATCAATGTTTTCATCCATCAHAAAVVGGCCACAGGAHTDATTCBCATCCACCAGATGADTTC

TGAAGTGTGAGTGTCTTAACCTTCAGTCAACCTGAAATAAACTAVTTHCTAATVTAACAGAA
TGACCTATCAGTGTCTTAACCTTCAGTCAACCTGAAATAAACTAVTTHCTAATVTAACAGAA
AGAAGTGTGAGTGTCTTAACCTTCAGTCAACCTGAAATAAACTAVTTHCTAATVTAACAGAA
TGAAGTGTGAGTGTCTTAACCTTCAGTCAACCTGAAATAGTACCAATTTCTAATATAACAGAA

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FIGURE 7

10 20 30 40 50 60 70 80
CCAAAGAAAAAGTGAATTGTCTATGCTTTATAGACTGTAAGAAGAGAACATCTCAGAAGTGGAGTCTTACCCCTGAAATCAAA
GAGTTTTATACCTCAATAGACT
10 20

90 100 110 120 130 140 150 160
GGATTAAAGAAAAAGTGAATTTTCTTCAGCAAGCTGTGAAACTAAATCCACAACCTTTGGAGACCCAGGAACACCCTCC
CTTACTAGTTTCTCTTTTTCAGGTGTGAAACTCAACCTTCAAAGACACTCTGTTCCATTCTGTGGACTAATAGGATCATC
30 40 50 60 70 80 90 100

170 180 190 200 210 220 230 240
AATCTCTGTGTGTTTGTAAACATCACTGGAGGGTCTTCTACGTGAGCAATTGGATTGTTCATCAGCCCTGCCTGTTTTCAC
TTTAGCATCTGCCGGGTGGATGCCATCCAGGCTTCTTTTCTACATCTCTGTTCTCGATTTTTGTGAGCCTAGGAGGTGCC
110 120 130 140 150 160 170 180

250 260 270 280 290 300 310 320
CTGGGAAGTGCCCTGGTCTTACTTGGGTCCAAATTGTGTGGCTTTCACTTTTGAACCTAAGCATCTGAAGCCATGGGCCACAC
TAAGCTCCATTGGCTCTAGATTCTGGCTTTCCCATCATGTTCTTCAAAGCATCTGAAGCTATGGCTTGCATTGTTCAGTT
190 200 210 220 230 240 250 260

330 340 350 360 370 380 390 400 410
ACGGAGGCAGGGAACATCACCATCCAAGTGTCCATACCTCAATTTCTTTTCAGCTCTTGGTCTGGCTGGTCTTTCTCACTTC
GATGCAGGATACCACTCTCTCAAGTTCCATGTCCAAGGCTCATTCTTCTTTGTGCTGCTGATTCTGTTTCAACAAGTG
270 280 290 300 310 320 330 340 350

420 430 440 450 460 470 480 490
TGTTTCAGGTGTTATCCACGTGACCAAGGAAGTGAAGAAGTGGCAACGCTGTCTGTGGTCACAAATGTTTCTGTGAAGAGC
TCTTCAGATGTTGATGAACAACGTGCAAGTCAGTGAAAGATAAGGTATTGCTGCCTTGCCGTTACAACCTCTCTCATGAAG
360 370 380 390 400 410 420 430

500 510 520 530 540 550 560 570
TGGCACAAACTCGCATCTACTGGCAAAAGGAGAAGAAATGGTGCTGACTATGATGTCTGGGGACATGAATATATGGCCCGA
ATGAGTCTGAAGACCGCAATCTACTGGCAAAACATGACAAAGTGGTGCTGTCTGTCTATTGCTGGGAAACTAAAGTGTGGCC
440 450 460 470 480 490 500 510

FIGURE 5-1

Contig
Murine B7-2
Porcine CD68(i)
Human B7.2

AATTCTGDCATAAATTTGACCTGCTCATCTAHACAAGGTTACCCAGAACCTAAGAAGATGTATD
AATTCTGTCTATAAATTTGACCTGCTCATCTACACAAGGCTACCCAGAACCCCGAGGATGTATA
AATTCTGGCATAAATTTGACCTGACGCTCTAAGCAAGGTCACCCGAAACCTAAGAAGATGTATT
AATGTGTACATAAATTTGACCTGCTCATCTATACACGGTTACCCAGAACCTAAGAAGATGAGTG

TTTGTCTAAVTACNAAGAATTCAACTAHTGAGTATGATGVTAAACATGCAGAAATCTCAAGATAA
TGTGTCTAAATACGAAGAATTCAACCACTGAGCATGATGCTGACATGAAGAAATCTCAAAATAA
TTCTGATAACT-----AATTCAACTAATGAGTATGGTGATAACATGCAGATATCAAGATAA
TTTTGTCTAAGAACCAAGAATTCAACTATCGAGTATGATGGTATTATGCAGAAATCTCAAGATAA

TGTCACAGAACTGTACAATGTHCTCCATCAGCBTGTCTCTTTTCATTCCCTGATGDTACGAGNNAT
CATCACGGAACCTCTACAATGTATCAATCAGGGTGTCTCTTCCCATCCCTCCCGAGACAA---AT
TGTCACAGAACTGTTCAGTATCTCCAACAGCCTCTCTCTTTTCATTCCCGGATGGTGTGTGGCAT
TGTCACAGAACTGTACGACGTTTCCATCAGCTTGTCTGTTTCATTCCCTGATGTTACGAGCAAT

ATGACCATCGTCTGTGTTCTGGAACTGAGNCAANCAAGACNCGCTTTTCTCCHHACCTTTCA
GTGAGCATCGTCTGTGTTCTGCAACTTGAGCCAAGCAAGACACTGCTTTTCTCCCTACCTTGTA
ATGACCGTTGTGTGTGTTCTGGAAACGGAGTCAATGAAGA-----TTTCTCTCAAACCTCTCA
ATGACCATCTTCTGTATTCTGGAACTGA-----CAAGACGCGGCTTTTATCTTCACTTTCT

ATATAGATCHAGAGBHHCCCTNNNCAACCTCCTNNCCCAGACCACATBCNNTGGATTACAGCTBT
ATATAGATGCAAAGCCACCTGTGCAACCCCTGTGCCAGACCACATCCTCTGGATTGCAGCTCT
ATTTCACTCAAGAGTTTCC-----ATCTCCTCAAACGTATGGAAG---GAGATTACAGCTTC
CTATAGAGCTTGAGGACCCT---CAGCCTCC---CCCAGACCACATTCCTTGGATTACAGCTGT

ACTTNNACAGTGGTCTVTTVTVTGTGTGATGGTGTCTTNTVTAATCTATGGAANNNAAGAAG
ACTTGTAAACAGTGGTCTGTTGTGTGTGGGATGGTGTCTTTGTAACACTAAGGAAA---AGGAAG
AGTT---ACTGTGGCCCTCTCTCTTGTGATGCTGCTC---ATCATTGTATG---TCACAAGAAG
ACTTCCAACAG---TTATTATATGTGTGATGGTTTTCTGTCTAATCTATGGAATGGAAGAAG

AAGAAGCAGCCTVGCATCTCTAATAAATGTGGNNNAACCAHCAAAATGGAGAGGGANGNGAGTG
AAGAAGCAGCCTGGCCCTCTAATGAATGTGGTGAAACCATCAAAATGAACAGGAAGGCGAGTG
CCGAATCAGCCTAGCAGGCCAGCAA-----CACAGCCTCTAAGTTAGAGCGGGA---TAGT-
AAGAAGCGGCCTCGCAACTCTTATAAATGTGG---AACCAACACAATGGAGAGGGAAGAGAGTG

AACANACTAAGAACAGAGAAAAANTCCATNNACCTGAAVGATCTGATGAAGCCCAGNGTGMINT
AACAACTAAGAACAGAGCAGAAGTCCAT-----GAACGATCTGATGATGCCCAGTGTGATGT
AAGC---CTG---ACAGAGAGA-----CTATCAACCTGAAGGAACT---TGAACCCCA-----
AACAGACCAAGAAAAGAGAAAAATCCATATACCTGAAAGATCTGATGAAGCCCAGCGTGTITT

TAANADTTNNAAGACAGCTTCANNNGACAAAAGTNNACANNTTTTTAADTNNAGAGTNAAGNN
TAATATTTTTAAGACAGCCTCAGATGACAACAGTACTACAGATTTTTAAGT-----
-----AATT-----GCTTCA-----GCAAAA-----CCAAATGCAGAGTGAAG--
TAAAGTTCGAAGACATCTTCATGCCACAAAAGTGATACATGTTTTTAATTAAAGAGTAAAGCC

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FIGURE 7-1

580 590 600 610 620 630 640 650
GTACAAGAACCGGACCATCTTTGATATCACTAATAACCTCTCCATGTGATCCTGGCTCTGGCGCCATCTGACGAGGGCACA
... ..
CGAGTATAAGAACCGGACTTTTATGACAACACTACCTACTCTCTTATCATCCTGGGCTGGTCTTTTACAGACCGGGGCACA
520 530 540 550 560 570 580 590

660 670 680 690 700 710 720 730
TACGAGTGTGTTGTCTGAAGTATGAAAAAGACGCTTTCAAGCGGGAACACCTGGCTGAAGTGACGTTATCAGTCAAAGCTG
... ..
TACAGCTGTGTCGTTCAAAGAAGGAAGAGGAACGTATGAAGTTAAACACTTGGCTTTAGTAAAGTTGTCCATCAAAGCTG
600 610 620 630 640 650 660 670

740 750 760 770 780 790 800 810 820
ACTTCCCTACACCTAGTATATCTGACTTTGAAATTCCAACTTCTAATATTAGAAGGATAATTGCTCAACCTCTGGAGGTTT
... ..
ACTTCTCTACCCCAACATAACTGAGTCTGGAAACCCATCTGCAGACACTAAAGGATTACCTGCTTTGCTTCCGGGGGTTT
680 690 700 710 720 730 740 750 760

830 840 850 860 870 880 890 900
TCCAGAGCCTCACCTCTCCTGGTTGGAAAATGGAGAAGAATTAAATGCCATCAACACAACAGTTTCCCAAGATCCTGAAACT
... ..
CCCAAAGCCTCGCTTCTCTTGGTTGGAAAATGGAAGAGAATTACCTGGCATCAATACGACAATTTCCAGGATCCTGAACTT
770 780 790 800 810 820 830 840

910 920 930 940 950 960 970 980
GAGCTCTATGCTGTAGCAGCAAACTGGATTTCATATGACAACCAACCAAGCTTCATGTGCTCATCAAGTATGGACATT
... ..
GAATTGTACACCATAGTAGCCAACTAGATTTCATACGACTCGCAACCAACCATTAAGTGTCTCATTAAATATGGAGATG
850 860 870 880 890 900 910 920

990 1000 1010 1020 1030 1040 1050 1060
TAAGAGTGAATCAGACCTTCAACTGGAATACAACCAAGCAAGAGCATTTTCTGATAACCTGCTCCCATCCTGGGCCATTAC
... ..
CTCAGTGTACAGGAGCTTACCTGGGAAAAACCCCAAGAGCCCTCCTGATAGCAAGAACACACTTGTGCTCTTTGGGGC
930 940 950 960 970 980 990 1000

1070 1080 1090 1100 1110 1120 1130 1140
CTTAATCTCAGTAAATGGAATTTTGTGATATGCTGCCTGACCTACTGCTTTGCCCCAAGATGCAGAGAGAGAAGGAGGAAT
... ..
AGGATTCGGCGCAGTAATAACAGTCGTCTCATCATCAATGCTTCTGTAAAGCACAGAAGCTGTTTCAGAAGA
1010 1020 1030 1040 1050 1060 1070 1080

FIGURE 7-2

1150 1160 1170 1180 1190 1200 1210 1220 1230
GAGAGATTGAGAAGGGAAAGTGTACGCCCTGTATAACAGTGTCCGCAGAAGCAAGGGGCTGAAAAGATCTGAAGGTAGCCCTC
... ..
AATGAGGCAAGCAGAGAAACAACAACAGCCTTACCTTCGGGCCTGAAGAAGCATTAGCTGAACAGACCGTCTTCCTTTAGT
1090 1100 1110 1120 1130 1140 1150 1160 1170

1240 1250 1260 1270 1280 1290 1300 1310
CGTCATCTCTTCTGGGATACATGGATCGTGGGGATCATGAGGCATTCTTCCCTTAACAAATTTAAGCTGTTTACCCACTAC
... ..
TCTTCTCTGTCATGTGGGATACATGGTATTATGTGGCTCATGAGGTACAATCTTTCTTTCAGCACCGTGTAGCTGATCTT
1180 1190 1200 1210 1220 1230 1240 1250

1320 1330 1340 1350 1360 1370 1380 1390
CTCACCTTCTTAAAAACCTCTTTTCAGATTAGCTGAACAGTTACAAGATGGCTGGCATCCCTCTCCTTTCTCCCATATGCA
... ..
TCGGACAACCTTGACACAAGATAGAGTTAACTGGGAAGAGAAAGCCTTGAATGAGGATTTCTTTCCATCAGGAAGCTACGGGC
1260 1270 1280 1290 1300 1310 1320 1330

1400 1410 1420 1430 1440 1450 1460 1470
ATTTGCTTAATGTAACCTCTTCTTTTGCCATGTTTCCATTCTGCCATCTTGAATTGTCTTGTTCAGCCAATTCATTATCTATT
... ..
AAGTTTGCTGGGCCTTTGATTGCTTGATGACTGAAGTGGAAAGGCTGAGCCCTAGTGGGTGGTGTAGCCCTGGGCAGGGG
1340 1350 1360 1370 1380 1390 1400 1410

1480 1490
AAACACTAATTTGAG
... ..
CAGGTGACCTGGGTGGTATAAGAAAAAGAGCTGTCACTAAAAGGAGAGGTGCCTAGTCTTACTGCAACTTGATATGTCATG
1420 1430 1440 1450 1460 1470 1480 1490

TTTGGTTGGTGTCTGTGGGAGGCCTGCCCTTTTCTGAAGAGAAGTGGTGGGAGAGTGCATGGGGTGGGGGCAGAGGAAAAGT
1500 1510 1520 1530 1540 1550 1560 1570 1580

GGGGGAGAGGGCCTGGGAGGAGAGGGAGGGGGACGGGGTGGGGTGGGGAAACTATGGTTGGGATGTAAAAACGGATA
1590 1600 1610 1620 1630 1640 1650 1660

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FIGURE 8a

10 20 30 40 50 60

Contig NNNNNNNNNNNNNNNNNNNNNNNNNNMTGCCNNCTGNNNNNNNNCTCGCCATGGTTCGTATTGCCTCTGCAG
Human CD40 GCCTCGCTCGGGCGCCCCAGTGCTCCTGCCGCGCTGGTCTCACCTCGCCATGGTTTCGTCTGCCTCTGCAG
Bovine CD40 -----CTCGCCATGGTTTCGTTCGCACTGCAG-----
Mouse CD40 -----TGCC--CTG-----CATGGTGTCTTCCTCGGCTG

70 80 90 100 110 120 130

Contig TCGCTCCTCTGGGGCTGCTTGCTGACCGCBGTCCATCCAGAACCACCCTGCDTGCAGAGAVAAACA
Human CD40 TCGCTCCTCTGGGGCTGCTTGCTGACCGCTGTCCATCCAGAACCACCCACTGCATGCAGAGAAAAACA
Bovine CD40 TGCTCTCTCTGGGGCTTCTTTCTGACCGCGTCCACTCAGAACCAGCCACTGCTTTGTGGAGAGAAGCA
Mouse CD40 TGCGCGCTATGGGGCTGCTTGTTGACAGCGGTCCATCTAGGGCAGTGTGTACGTGCAGTGACAAAACA

140 150 160 170 180 190 200

Contig GTACCTAIVTAACAGTCAGTGCTGTGATTGTGTGCCAGCCAGGACAGAAACTGGTGAGCGACTGCACAG
Human CD40 GTACCTAATAAACAGTCAGTGCTGTTCPTTGTGTGCCAGCCAGGACAGAAACTGGTGAGTGACTGCACAG
Bovine CD40 ATACCCAGTGAACAGTCTTTTCTGTGATTGTGTCCC CGCGGACAGAAACTGGTGAACGACTGCACAG
Mouse CD40 GTACCTCCACGATGGCCAGTGCTGTGATTGTGTGCCAGCCAGGAAGCCGACTGACAAGCCACTGCACAG

210 220 230 240 250 260 270

Contig AGBTCAVBAAAACVGAATGCCABCCHTGCGGTDAAGGCGAATCTCTTAGCCACCTGGAAACAGAGAGAHA
Human CD40 AGTTCACTGA AACCGAATGCCTTCTTTGCGGTGAAGCGAATCTCTAGACACCTTGAACAGAGAGACA
Bovine CD40 AGGTCAGCAAACAGAAATGCCAGTCTTCGGGTAAAGGCGAATCTCTTGCCACCTTGAACAGAGAGAAA
Mouse CD40 CTCTTGAGAAGACCCAATGCCACCCATGTGACTCAGGCGAATCTCAGCCCAGTGAACAGGGAGATT

280 290 300 310 320 330 340

Contig CACTGTCA CCAGCACAGATACTGCGACCCCAACCTAGGGCTTCGGGTCCAGAAGGAGGGCACCTCAGA
Human CD40 CACTGCCACCAGCACAAATACTGCGACCCCAACCTAGGGCTTCGGGTCCAGCAGAAGGGCACCTCAGA
Bovine CD40 TACTGTCA CAGCACAGATACTGCAACCCCAACCTAGGGCTCCGGATCCAGAGCGAGGGTACCTTGAA
Mouse CD40 CGCTGTCA CCAGCACAGACTGTGAACCCCAATCAAGGGCTTCGGGTTAAGAAGGAGGGCACCCGAGA

350 360 370 380 390 400

Contig AACAGACACCATCTGTACCTGTGTAVGAAGGCCAACACTGTACCAGTVAGGCCCTGCGAGAGHITGTGBCB
Human CD40 AACAGACACCATCTGCACCTGTGAAGAAGGCTGGCACTGTACGAGTGAGGCCTGTGAGAGCTGTGTCC
Bovine CD40 TACAGACACCATTTGTGTATGTGTGCGAAGGCCAACACTGTACCAGTCAACCTGCGAAAGTTGCACGC
Mouse CD40 ATCAGACACTGTCTGTACCTGTAAAGGAAGGACAACACTGCACCAGCAAGGATTGCGAGGCATGTGCTC

410 420 430 440 450 460 470

Contig HGCACAGCTCTVTGTHTCCTTGGCTTTGGGGTCAAGCAGATGCGTACAGGGVVTTCTGATACCGTCTGT
Human CD40 TGCACCGCTCATGCTCGCCCGGCTTTGGGGTCAAGCAGATTGCTACAGGGGVTTCTGATACCATCTGC
Bovine CD40 CCCACAGCTTGTGTCTCCCTGGCTTCGGGGTCAAGCAGATCGCTACAGGGGVTTTGGATACCGTCTGT
Mouse CD40 AGCACAGCCCTGTATCCCTGGCTTTGGAGTTATGGAGATGGCCACTGAGACCACTGATACCGTCTGT

480 490 500 510 520 530 540

Contig GADCCCTGCCCAGTCGGCTTCTTCTCCAATGTGTTCATCTGCTTTGAAAAGTGTACCCCTTGGACAAG
Human CD40 GAGCCCTGCCCAGTCGGCTTCTTCTCCAATGTGTTCATCTGCTTTGAAAAGTGTACCCCTTGGACAAG
Bovine CD40 GAACCTCGCCGCTCGGCTTCTTCTCCAACGTGTATCTGTTTTGAAAAGTGTACCCGTTGGACAAG
Mouse CD40 CATCCCTGCCCAGTCGGCTTCTTCTCCAATCAGTCATCACTTTTGA AAAAGTGTATCCCTGGACAAG

550 560 570 580 590 600 610

Contig CTGTGAGAVHAAGACCTGGTGGTVC A ACAGGHAGGVACGAACAAGACTGATGTTGTCTGTGGTTTCC
Human CD40 CTGTGAGACCAAAGACCTGGTGTGTCAACAGGCAGGCACAAACAAGACTGATGTTGTCTGTGGTCCCC
Bovine CD40 CTGCGAGAGAAAAGGCCTGGTGGACAACACGTGGGACGAACAAGACAGATGTTGTCTCGGGTTTCC
Mouse CD40 CTGTGAGGATAAGAACTTGGAGGTCTACAGAAAAGAACAGAGTCAGACTAATGTTCATCTGTGGTTTAA

620 630 640 650 660 670 680

FIGURE 8a-1

Contig AGDVTGCGGATGAGAGCCCTGGTGGTGAATCCCGTCATGATGGGVATCCTGTTTGCCATCCTCTTGGTG
Human CD40 AGGATCGGCTGAGAGCCCTGGTGGTGAATCCCGTCATCTTCGGGATCCTGTTTGCCATCCTCTTGGTG
Bovine CD40 AGAGTCGGATGAGGACCCCTGGTGGTGAATCCCGTCACGATGGGAGTCTTGTGTTGCTGTCTCTTGGTA
Mouse CD40 AGTCCCGGATGCGAGCCCTGCTGGTCAATCCTGTCTGTGATGGGCATCCTCATCACCATTTTCGGGGTG

690 700 710 720 730 740

Contig TTTGTCTDTATCAAAAAGGTGGCCAAAGAAGCCAACVGATAANNNGGCCCTVCAACCTANGGCTNNANG
Human CD40 CTGGTCTTTATCAAAAAGGTGGCCAAAGAAGCCAACCAATAA---GGCCCCCAACCCA-----A
Bovine CD40 TCTGCCTGTATCAGGAACATAACCAAGAAGC-GGCAGCTAA---GGCCCTGCACCCCTATGGCTGAAAG
Mouse CD40 TTTCTCTATATCAAAAAGGTGGTCAAGAAACCAAGGATAATGAGATGTTACCCCTGCGGCTCGACG

750 760 770 780 790 800 810

Contig GCAGGATCCCCAGGAGATGANTNNYCCNGAVGATTTTCCCGGCCCAACACCGCTGCTCCAGTGCAGG
Human CD40 GCAGGAACCCCAGGAGATCAATTTTCCCGACGATCTTCTGCTGCTCCAACTGCTGCTCCAGTGCAGG
Bovine CD40 GCAGGATCCCGTGGAGACGATTTGATCCCGGAGGATTTTCCCGGCCCAAC-CCGCTCTCCGGTGCAAG
Mouse CD40 GCAAGATCCCCAGGAGATG-----GAAGATTATCCCGGTCATAACACCGCTGCTCCAGTGCAGG

820 830 840 850 860 870 880

Contig AGACTTTACACGGGTGTGAGCCCGTACCCAGGAGGATGGCAAAGAGAGTCCGATCTCAGTGCAGGAG
Human CD40 AGACTTTACATGGATGCCAACCGGTACCCAGGAGGATGGCAAAGAGAGTCCGATCTCAGTGCAGGAG
Bovine CD40 AGACCTTATGCTGGTGTGAGCCCGTCCGCCAGGAGGACGGCAAAG
Mouse CD40 AGACACTGCACGGGTGTGAGCCGTGTACACAGGAGGATGGTAAAGAGAGTCCGATCTCAGTGCAGGAG

890 900 910 920 930 940 950

Contig CGGCAGGTGACAGACAGCATAGCCTTGAGGCCCCCTGGTCTGMAACCTGGAACVGCCTTYRGRRGYGATG
Human CD40 -----AGACAG-----TGAGGC-----TGCAACC-----ACC-----CAGGAGTG-TG
Mouse CD40 CGGCAGGTGACAGACAGCATAGCCTTGAGGCCCCCTGGTCTGAACCTGGAACVGCCTTTGGAGGCGATG

960 970 980 990 1000 1010 1020

Contig# 1 GCYRCTGTGCTGACCTTTGAAGTTTGAGRTGRGCCAARACAGAGCCCAAGTGCAGYTTRCYCTCATGCCT
Human CD40 GCCAC-----GTGGGC-----AAACAG-----GCAGTTGGCC-----
Mouse CD40 GCTGCTTGTGACCTTTGAAGTTTGAGATGAGCCAAGACAGAGCCCAAGTGCAGCTAACTCTCATGCCT

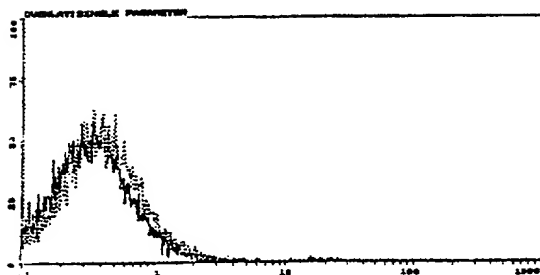
Figure 8b

	10	20	30	40	50	60
Contig
bovine CD40 protein	MVRLPLQCLFWGFFLTAVHSE	PATACGEKQYFVNSLCCDLC	PPGQKLVNDCTEVSKTECQ			
human CD40 protein	MVRLPLQCVLWGCILLTAVH	PEPPTACREKQYLINSQCC	SLCQPGQKLVSDCTEFTET	ECLE		
murine CD40 protein	MVSLPRLCALWGCILLTAVH	LGCQVTCSDKQYLHDGQC	CDLCQPGSRLTSHCTALEK	TQCH		
	70	80	90	100	110	120
Contig
bovine CD40 protein	SCGKGFEFLSTWNREKYCHE	HRYPNPNLGLRIQSEGT	LNTDTICVCVEGQHCTSH	TCE	SC	T
human CD40 protein	PCGESEFLDTYNRETHCHQ	HKYCDPNLGLRVQQKGT	SETDTICTCEGWHCTSE	AC	ESC	V
murine CD40 protein	PCDSGEFSAQWNREIRCHQ	HRHCEPNQGLRVKKEGT	AESDVTCTCKEGQHCT	SKD	CE	ACA
	130	140	150	160	170	180
Contig
bovine CD40 protein	PHSLCLPGFGVKQIATGL	LDTVCEPCPLGFFSNV	SSAFEKCHRWTSCERK	GLVEQH	VG	TN
human CD40 protein	LHRSCSPGFGVKQIATG	VSDTICEPCPVGFFSN	VSSAFEKCHPWTSCE	TKDLV	VQ	AG
murine CD40 protein	QHTPCIPGFGVMEMATET	ITDTVCHPCPVGFFSN	QSSLFKCYPWTSC	EDKNLE	VLQ	KG
	190	200	210	220	230	240
Contig
bovine CD40 protein	KTDVVCGFQSRMRTL	VVIFVIMGVLFVAVLL	VSACIRNITTK-----	RQLRP	CTL	
human CD40 protein	KTDVVCGPQDRLRAL	VVIFIIFGILFAILL	VLVFIKKVAKKPTNK	APHP----	KQEPQ	EI
murine CD40 protein	QTNVICGLKSRMRALL	VIPVVMGILITIFG	VFLYIKKVVKPKDNE	MLPPAARRQ	DPQ	EM
	250	260	270	280		
Contig	
bovine CD40 protein	WLKGRIPWRL---	IRRIFFA--PTRLS	GARDMLV	SAGRP	GG	RQ
human CD40 protein	NFPDDLPGSNTA	APVQETLHGCQ	PVTQEDG	KESRISV	QERQ	
murine CD40 protein	---EDYPGHNTA	APVQETLHGCQ	PVTQEDG	KESRISV	QERQ	VTD

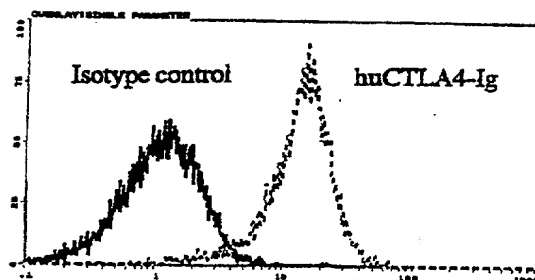
FIGURE 9

A

Non-transfected control cells

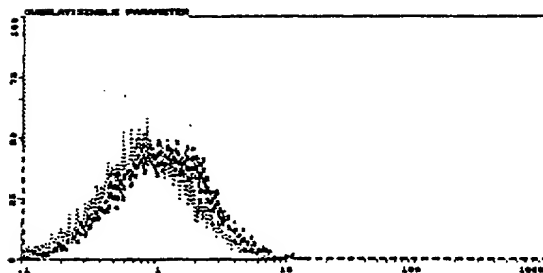


Transfected cells



B

Non-transfected control cells



Transfected cells

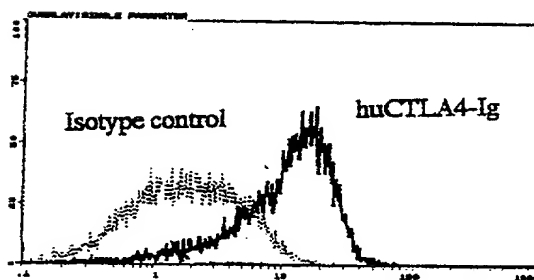
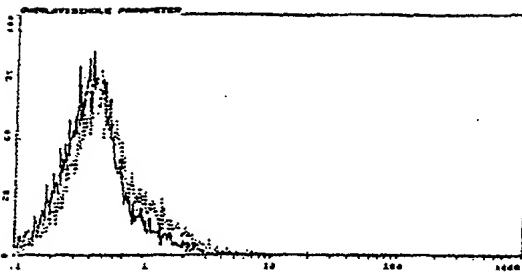
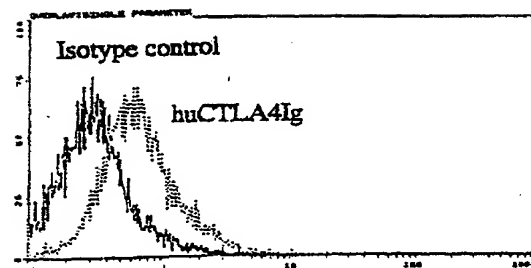


FIGURE 10

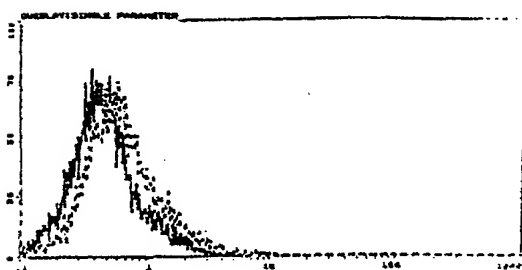
Non-transfected control cells



Transfected cells



Non-transfected control cells



Transfected cells

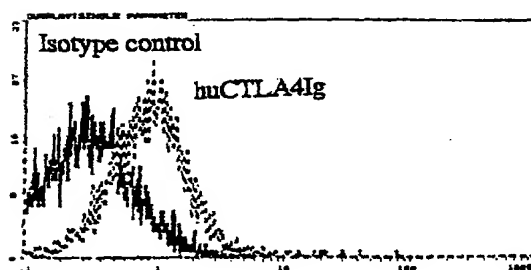
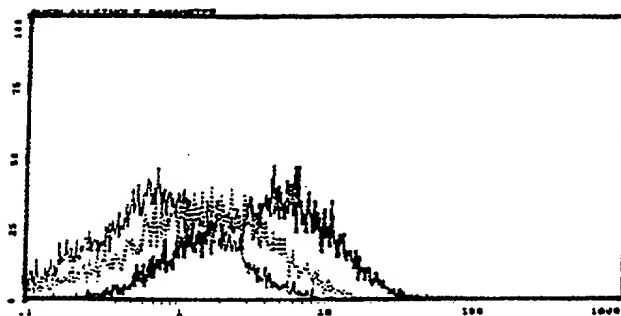


FIGURE 11

A



B

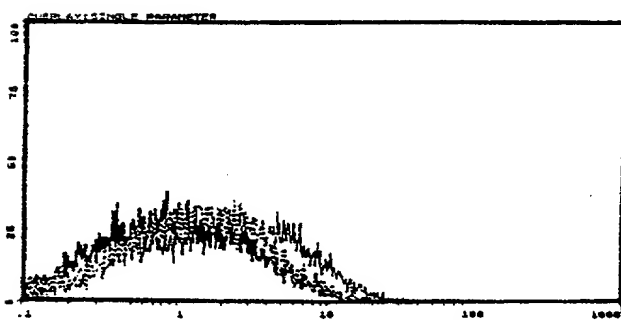
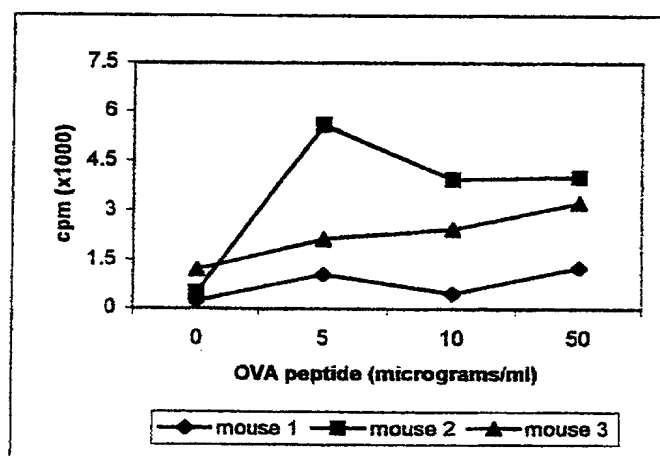
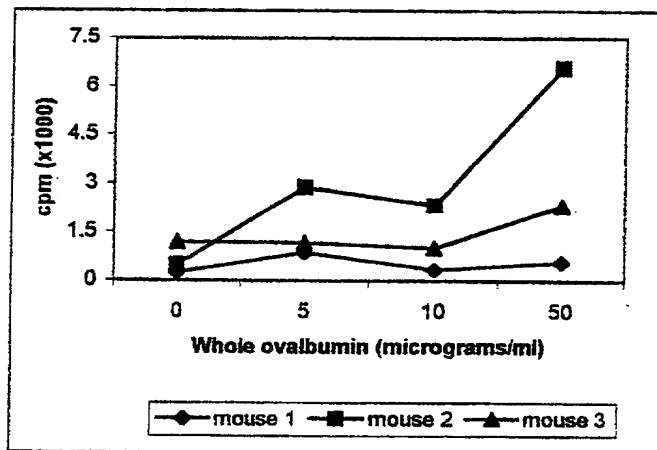


FIGURE 12

1 MGLSNILFVM VLLLSGAASL KSQAYFNETG ELPCHFTNSQ
41 NLSLDELVIF WQDQDNLVLY ELYRGQEKPH NVNSKYMGR
81 SFDQATWTLR LHNVQIKDKG SYQCFIHHKG PHGLVPIHQM
121 SSDLSLLANF SQPEINLLTN HTENSVINLT CSSTQGYPEP
161 QRMYYMLLNTK NSTTEHDADM KKSQNNITEL YNVSIRVSLP
201 IPPETNVSIV CVLQLEPSKT LLFSLPCNID AKPPVQPPVP
241 DHILWIAALL VTVVVVCGMV SFVTLRKRKK KQPGPSNECG
281 ETIKMNRKAS EQTKNRAEVH ERSDDAQCDV NILKTASDDN
321 STTDF•LKSK L

FIGURE 13



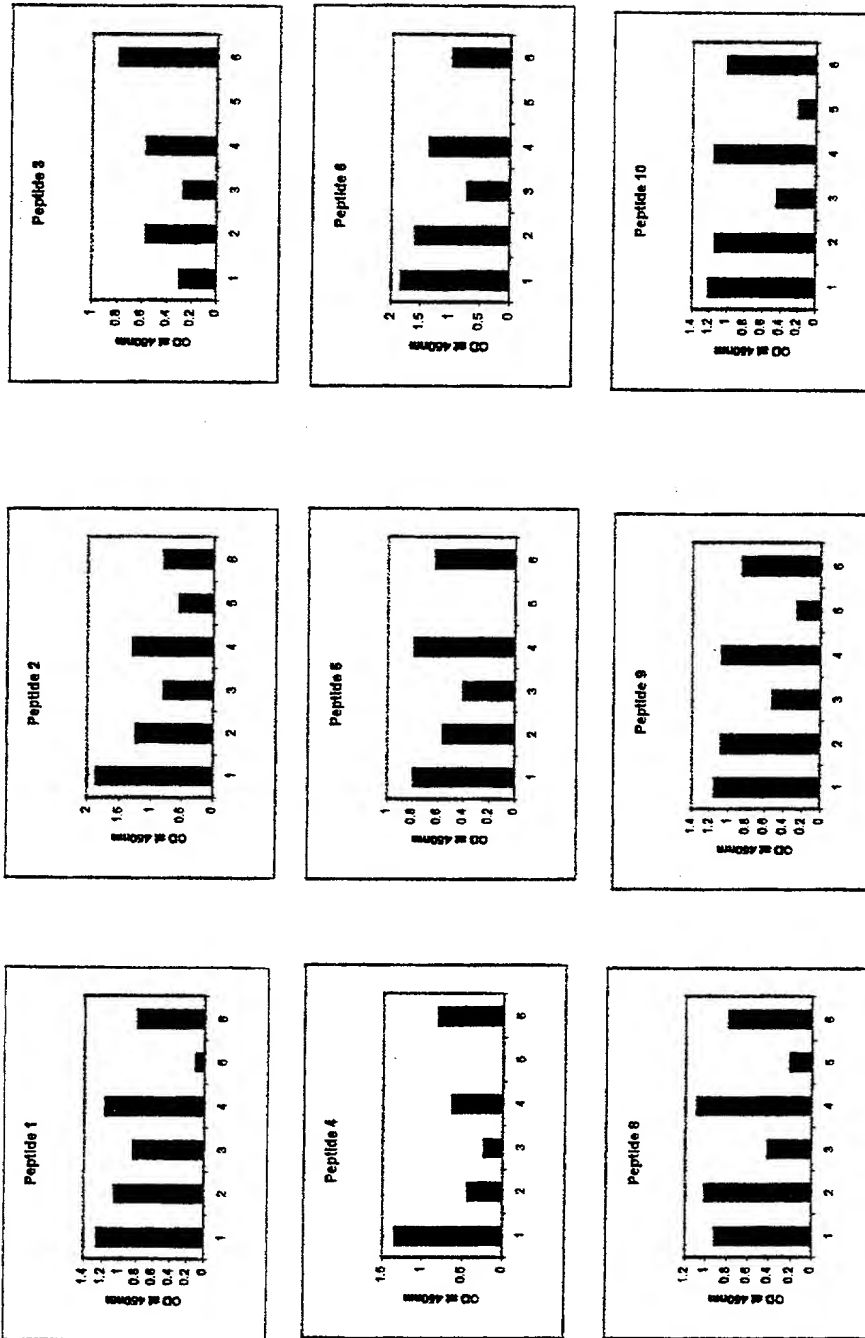


Figure 14a

FIGURE 14b

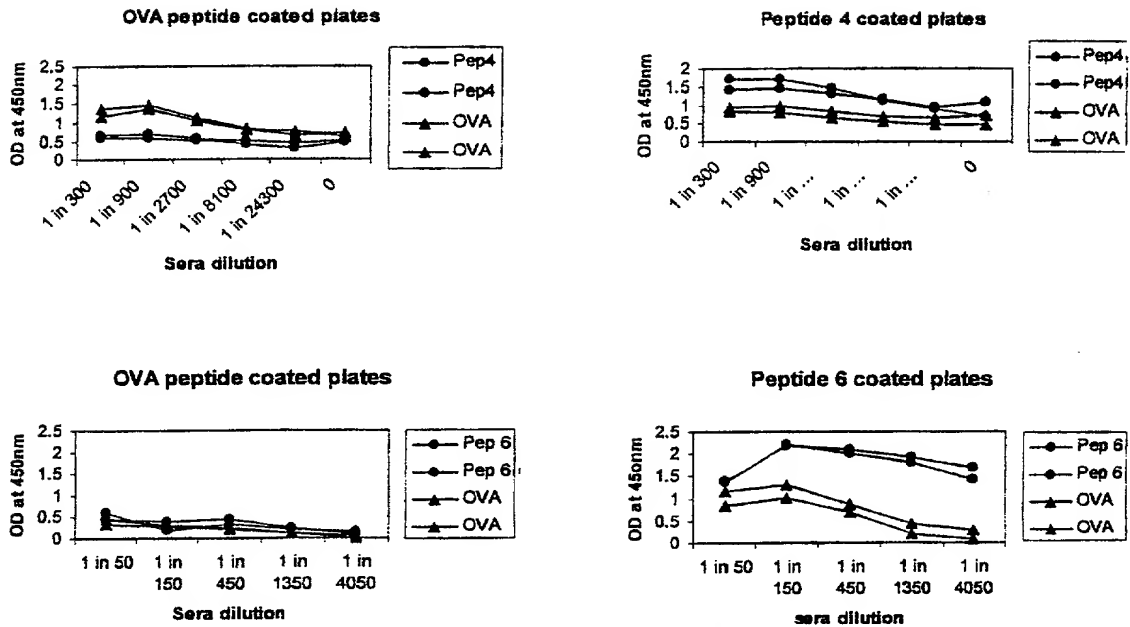


FIGURE 15a

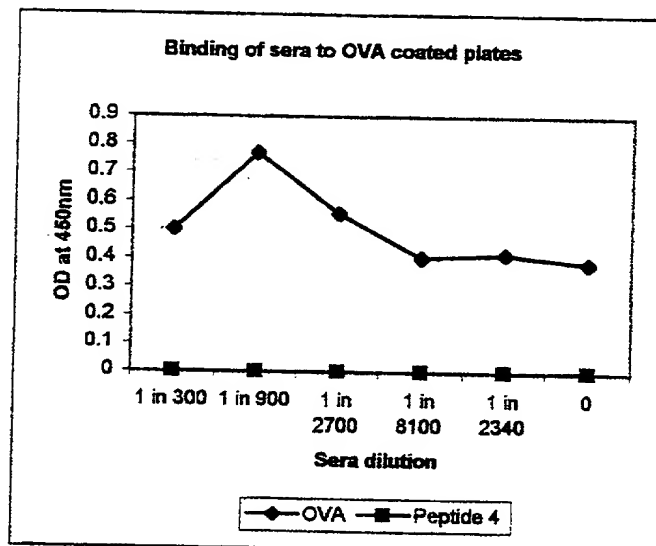
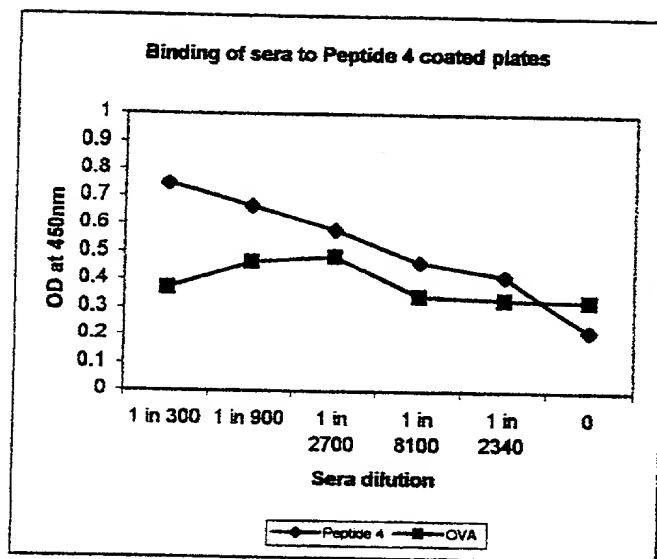


FIGURE 15b

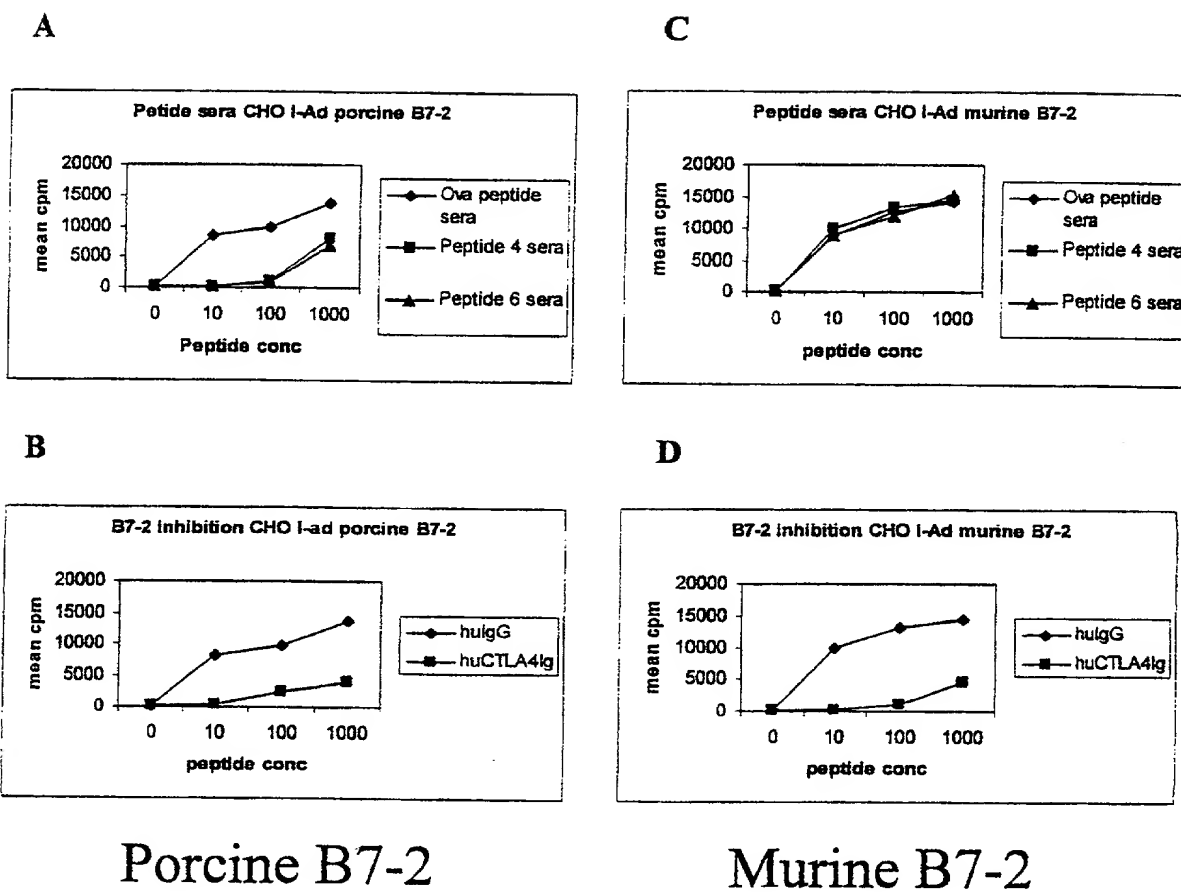


Figure 1b

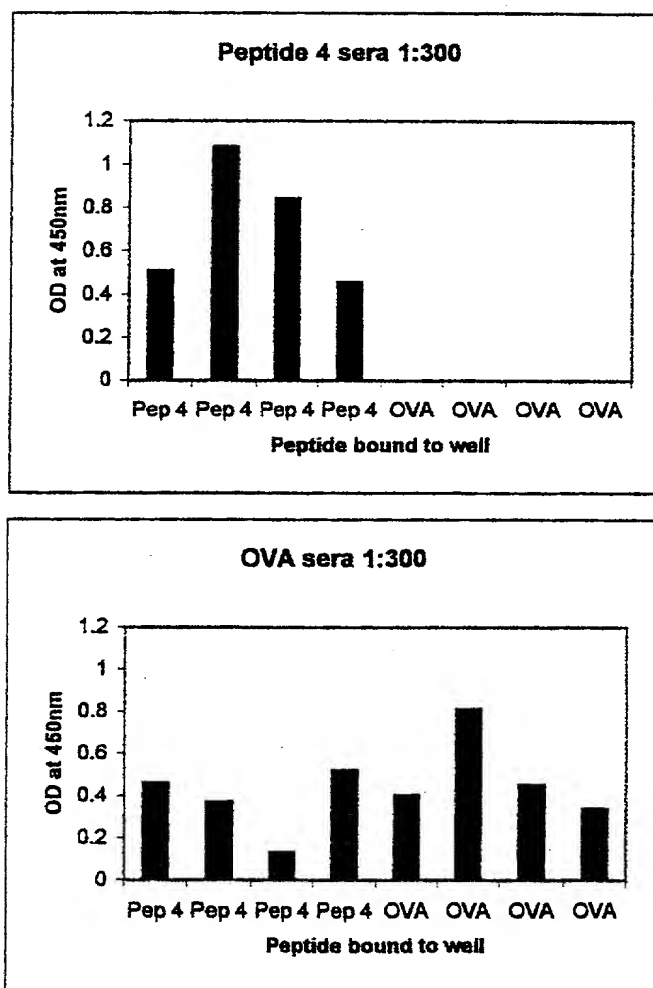
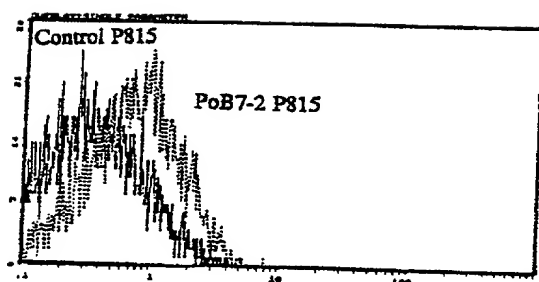
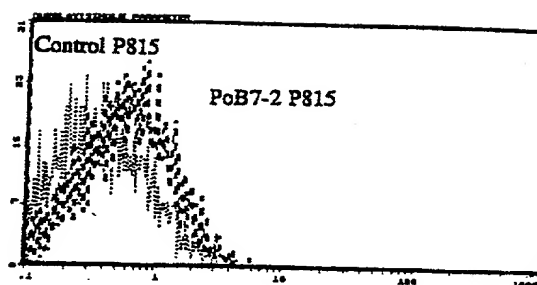


FIGURE 17a

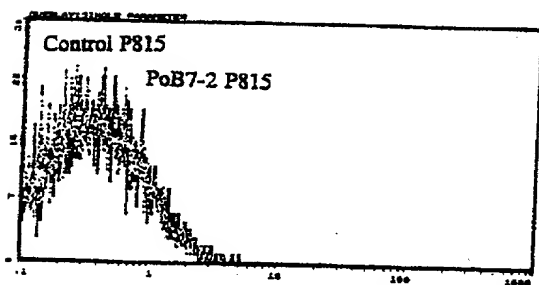
A



B



C



D

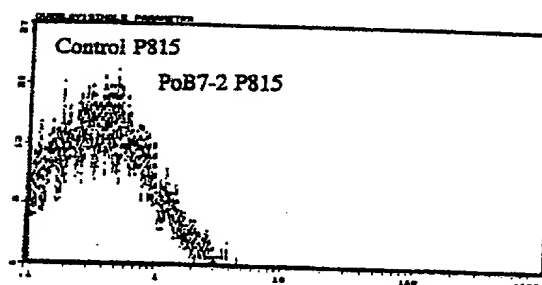


FIGURE 17b

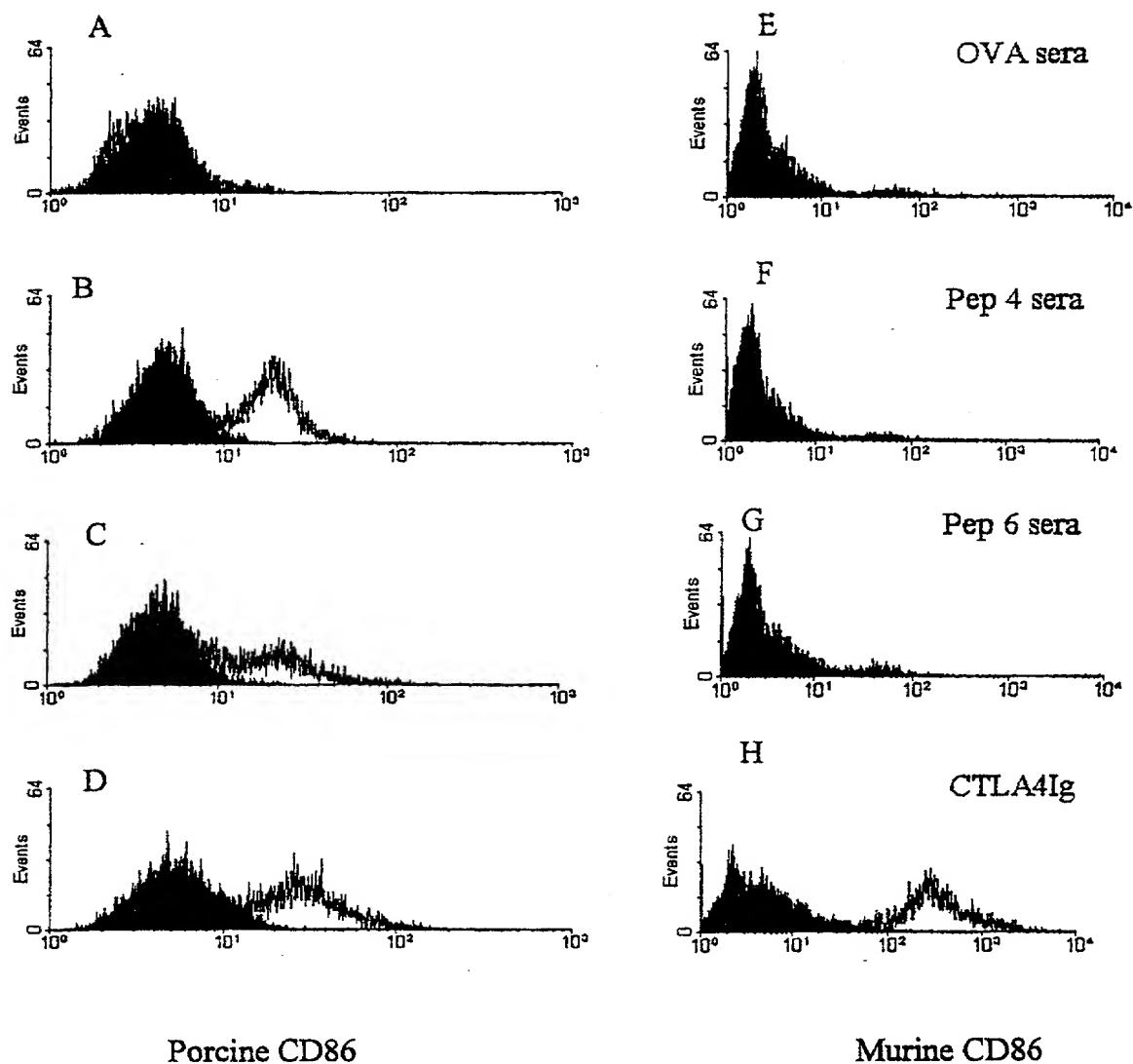


Figure 18

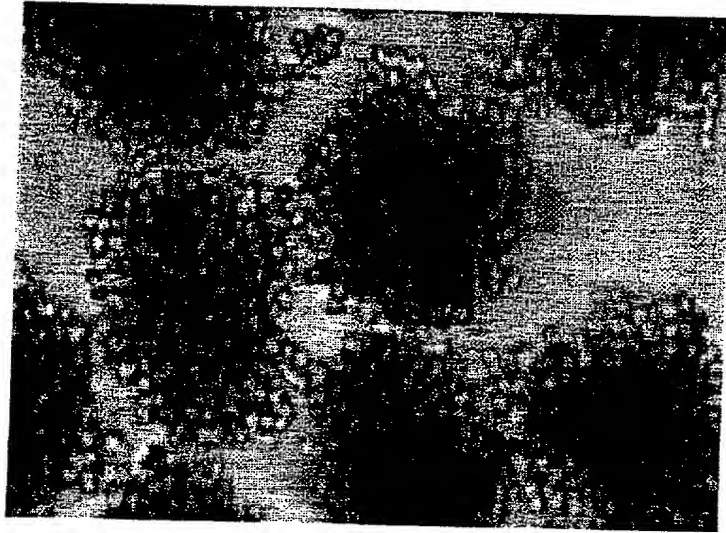


FIGURE 19

Day 1: Immunisation of C57BL-6 mice with whole ovalbumin (50 micrograms) in Complete freunds adjuvant (CFA)



Day 14: First immunisation with chimeric peptide (100 micrograms) i.v.

Day 21: Second immunisation with chimeric peptide (100 micrograms) i.v.

Day 28: Third immunisation with chimeric peptide (100 micrograms) i.v.



Day 32: Mice rendered diabetic by injection of streptozotocin i.p.

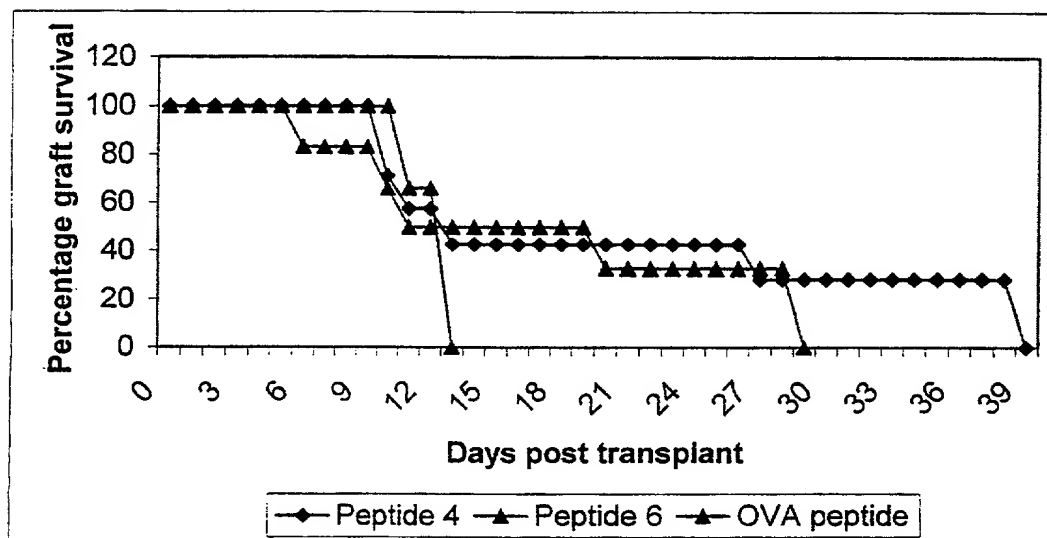


Day 36 : Transplantation of 1000 porcine pancreatic islets under the kidney capsule of diabetic mice



Day 37 onwards : Survival of islets assessed by measuring blood glucose levels

Figure 20



poCD40protein(top), human CD40 protein(bottom)

10 20 30 40 50 60 70 80
MVRLPLQCLLWGCFLLTAVHPEPPTSCKENQYPTNSRCCNLCPGQKLVNHCTEVTETECLPCSSSEFLATWNREKHCHQHKY
.....
MVRLPLQCVLWGCLLTAHVHPEPPTACREKQYLINSQCCSLCQPGQKLVSDCTEFTETECLPCGESEFLDTWNRETHCHQHKY
10 20 30 40 50 60 70 80

90 100 110 120 130 140 150 160
CDPNLGLQVQREGTSKTDITTCVCSEGHCTNSACESCTLHSLCFGLGVKQIMATEVSDITICEPCFVGFFSNVSSASEKCPW
.....
CDPNLGLRVQQKGTSETDTICTCEEGWHTCTSEACESCVLHRSCSPGFGVKQIATGVSDITICEPCFVGFFSNVSSAFKCHPW
90 100 110 120 130 140 150 160

170 180 190 200 210 220 230 240
TSCESKGLVEQRAGTINKTIDVVGCFQSRMRALVVIPTILGILFAVLLVFLCIRKVTKEQETKALHPKTERQDFVETIDLEDFP
.....
TSCETKDLVVQAGTINKTIDVVGCPQDRLRALVVIPIIFGILFAILLVLVFIKKVAKKPTINKAPHPKQEPQEIINFDDLPGSN
170 180 190 200 210 220 230 240

250 260 270
DSTAPVQETLHWCPVTQEDGKESRISVQERQ
.....
TA-APVQETLHGCQPVVTQEDGKESRISVQERQ
250 260 270

Figure 22

1 MVRLPLQCLL WGCFLTAVHP EPPTSCKENQ YPTNSRCCNL
41 CPPGQKLVNH CTEVTETECL PCSSEFLAT WNREKHCHQH
81 KYCDPNLGLQ VQREGTSKTD TTCVCSEGGH CTNSACESCT
121 LHSLCFPGLG VKQMATEVSD TICEPCPVGF FSNVSSASEK
161 CQPWTSCESK GLVEQRAGTN KTDVVCGFQS RMRALVVIPI
201 TLGILFAVLL VFLCIRKVTK EQETKALHPK TERQDPVETI
241 DLEDFPDSTA PVQETLHWCQ PVTQEDGKES RISVQERQ

Figure 23

pig VCAM peptide copy(top), human VCAM peptide copy(bottom)

-20 -10 10 20 30 40 50 60
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
IVVIFGASNILMMVFAVSQNVKVEIFPEDKMLAQIGDSASLTCSAPDCSSLSFSWRTQIDSPLNGKVKTNGTRSTLVMNFV
.....
MVLILGASNILWIMFAASQAFKIETTPESRYLAQIGDSVSLTCSITGCESP-FFSWRTQIDSPLNGKVINEGTITSTLTMMNFV
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
10 20 30 40 50 60 70 80

70 80 90 100 110 120 130 140
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
SFENEHSYLCTVSCGNLKGGERGIQVEIYSFPKDPPEIHWSLPEVGKPVTVRCLVPDVYPVEKLEIELLKDNHSMVSONFLEL
.....
SFGNEHSYLCTATCESRKLEKGIQVEIYSFPKDPPEIHLGPLEAGKPITVKCSVADVYPFDRLEIDLKGDHLMKSQEFLEL
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
90 100 110 120 130 140 150 160

150 160 170 180 190 200 210 220
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
IDIKSKETKSLEFTFTPTTEEDIGKALVCOATLIIDGQPSVKTTPEKM---QVYISPKDPVISVNPSTSLQEGDSMMMTCTSE
.....
ADRSKLETKSLEVTFTFPVIEDIGKVLVCRAKLHIDEMDSVPTVRQAVKELQVYISPKNTVISVNPSTKLQEGGSSVTMTCSSE
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
170 180 190 200 210 220 230 240

230 240 250 260 270 280
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
GLPAPQISWSKKLNGDQQLSGNATLTLLIAMRMEDSGIYVCEGVNPGTNRKEVELIVQ-----
.....
GLPAPEIFWSKKLDNGNLQHLSGNATLTLLIAMRMEDSGIYVCEGVNLIGKNRKEVELIVQEKPFIVEISPGPRIAAQIGDSV
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
250 260 270 280 290 300 310 320

MLTCSVMGCESPSFSWRTQIDSPLSGKVRSEGTNSTLTLSPVSFENEHSYLCTVTCGHKKLEKGIQGEIYSFPDRDPEIEMSG
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
330 340 350 360 370 380 390 400

GLVNGSSCTVSCVKVPSVYPLDRLEIELLKGETILENIEFLEDITOMKSLENKSLEMTFIPTIETDTGKALVCOAKLHIDMEFE
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
410 420 430 440 450 460 470 480 490

290 300 310 320 330 340 350
|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
-----VAPRDTTISVNPSSTLEEGSSVNMTCSDDGFPAPKILWSKKLRDGNLEPLSENTTLTSTKRMEDSGIY
.....

FIGURE 23-1

360 370 380 390 400 410 420 430
VCEGINQAGINRKEVELIIQAAPKDLQLTAFPSSESVKEGDTVLIISCTCGNVPPTLIILKKKAETGDTV LKSTDGAYTTIHRAR
.....
LCEGINQAGRSRKEVELIIQVTPKDIKLTAFPSSESVKEGDTVLIISCTCGNVPETWIIILKKKAETGDTV LKSIDGAYTIRKAQ
580 590 600 610 620 630 640 650

440 450 460 470 480 490 500 510
LADAGVYECESKNEIGLQLRSTTL DVKGRESNKDYFSSELLVLYCASSLIIPAIGVLIYFARKANMRGYSYSLVDAQSKV
.....
LKDAGVYECESKNKVGSQLRSLTLDVQGRENNKDYFSPELLVLYFASSLIIPAIGMIYFARKANMRGYSYSLVEAQSKV
660 670 680 690 700 710 720 730

Figure 24

↓ (signal sequence)

IVVIFGASNI LWMVFAVSQN VKVEIFPEDK MIAQIGDSAS
LTCSAPDCES SLSFSWRTQI DSPLNGKVKT NGTRSTLVMN
PVSFENEHSY LCTVSCGNLK GERGIQVEIY SFPKDPEIHW
SSLPEVGKPV TVRCLVPDVY PVEKLEIELL KDNHSMVSQN
FLELIDIKSK ETKSLEFTFT PTEEDIGKAI VCQATLIIDG
QPSVKTTP EK MQVYISPKDP VISVNPSTSL QEGDSMMMT C
TSEGLPAPQI SWSKKLDNGD QQLLSGNATL TLIAMRMEDS
GIYVCEGVNP VG TNRKEVEL TVQVAPRDTT ISVNPSSSTLE
EGSSVNMTC S SDGFPA PKIL WSKKLRDGNL EPLSENTTLT
LTSTKMEDSG IYVCEGINQA GINRKEVELI IQAAPKDLQL
TAFPSESVKE GDTVIIISCTC GNVPPTLIIL KKKAETGDTV
LKSTDGAYTI HRARLADAGV YECESKNEIG LQLRSITLDV
KGRESNKDYF SSELLVLYCA SSLIIPAIGV IIYFARKANM
RGSYSLVDAQ KSKV•

FIGURE 25

translated po B7-2 Maher(top), human B7-2 translated(bottom)

```

      10      20      30      40      50      60      70      80
MGLSNILFVMVLLLSGAASLKSQAYFNETGELPCHFTNSQNLSDLVIFWQDQDNLVLYELRGQEKPHNVNSKYMGRITSF
.....
MGLSNILFVMAFLLSGAAPLKIQAYFNETADLPQFANSQNSLSLVVFWQDQENLVINEVYLGKEKFDVHSKYMGRITSF
      10      20      30      40      50      60      70      80

      90     100     110     120     130     140     150     160
DQATWTLRLHNVQIKDKGSYQCFIHHKGPHGLVPFHQMSSDLLANFSQPEINLLINHTENSVINLTCSSIQGYPEPQRMV
.....
DSDSWTLRLHNLQIKDKGLYQCIHHKKPTGMIRIHQMNSLSVLNFSQPEIVPISNITENVYINLTCSSIHGYPEPKIMS
      90     100     110     120     130     140     150     160

      170     180     190     200     210     220     230     240
MLLNTKNSTTEHDADMKKSQNNITELYNVSIKRVSLPIPETNVSIVCVLQLEPSKTLFLSLPCNIDAKPPVQPPVDPHILWI
.....
VLLRTKNSTLEYDGIQKSQDNVTELYDVSIKSVSFPDVTSNMIFCILETDKTRILLSSPFSIELEDPPPDHIPWITAV
      170     180     190     200     210     220     230     240

      250     260     270     280     290     300     310     320
AALLVTVVVVCGMVSVFTLRKRKKKQPGPSNECGETIKMNRKASEQTKNRAEVHERSDDAQCDVNILKTASDDNSTTDF
.....
LPVILICVMVFCLILWKKKKRPRNSYKCGTNIMEREESQTKKREKIHIPERSDEAQRVFKSSRTSSCDKSDTCF
      250     260     270     280     290     300     310     320

```

FIGURE 26

1 MGLSNILFVM VLLLSGAASL KSQAYFNETG ELPCHFTNSQ
41 NLSLDELVIF WQDQDNLVLY ELYRGQEKPH NVNSKYMGRT
81 SFDQATWTLR LHNVOIKDKG SYQCFIHHKG PHGLVPIHQM
121 SSDLSLLANF SQPEINLLTN HTENSVINLT CSSTQGYPEP
161 QRMVYMLLNTK NSTTEHDADM KKSQNNITEL YNVSIRVSLP
201 IPPETNVSIV CVLQLEPSKT LLFSLPCNID AKPPVQPPVP
241 DHILWIAALL VTVVVVCGMV SFVTLRKRKK KQGPSNECG
281 ETIKMNRKAS EQTKNRAEVH ERSDDAQCDV NILKTASDDN
321 STTDF

SEQUENCE LISTING

<110> ML Laboratories PLC

<120> Immunosuppression

<130> P15700WO

<140> PCT/GB99/04200

<141> 1999-12-17

<150> 9827921.9

<151> 1998-12-19

<150> 9925015.1

<151> 1999-10-23

<160> 39

<170> PatentIn Ver. 2.1

<210> 1

<211> 288

<212> PRT

<213> Homo sapiens

<400> 1

Met	Gly	His	Thr	Arg	Arg	Gln	Gly	Thr	Ser	Pro	Ser	Lys	Cys	Pro	Tyr
1				5					10					15	

Leu	Asn	Phe	Phe	Gln	Leu	Leu	Val	Leu	Ala	Gly	Leu	Ser	His	Phe	Cys
			20					25						30	

Ser	Gly	Val	Ile	His	Val	Thr	Lys	Glu	Val	Lys	Glu	Val	Ala	Thr	Leu
		35					40					45			

Ser	Cys	Gly	His	Asn	Val	Ser	Val	Glu	Glu	Leu	Ala	Gln	Thr	Arg	Ile
	50					55					60				

Tyr	Trp	Gln	Lys	Glu	Lys	Lys	Met	Val	Leu	Thr	Met	Met	Ser	Gly	Asp
65					70					75					80

Met	Asn	Ile	Trp	Pro	Glu	Tyr	Lys	Asn	Arg	Thr	Ile	Phe	Asp	Ile	Thr
				85					90					95	

Asn	Asn	Leu	Ser	Ile	Val	Ile	Leu	Ala	Leu	Arg	Pro	Ser	Asp	Glu	Gly
			100					105						110	

Thr	Tyr	Glu	Cys	Val	Val	Leu	Lys	Tyr	Glu	Lys	Asp	Ala	Phe	Lys	Arg
		115					120					125			

Glu	His	Leu	Ala	Glu	Val	Thr	Leu	Ser	Val	Lys	Ala	Asp	Phe	Pro	Thr
		130				135					140				

Pro	Ser	Ile	Ser	Asp	Phe	Glu	Ile	Pro	Thr	Ser	Asn	Ile	Arg	Arg	Ile
145					150					155					160

Ile	Cys	Ser	Thr	Ser	Gly	Gly	Phe	Pro	Glu	Pro	His	Leu	Ser	Trp	Leu
				165					170					175	

Glu	Asn	Gly	Glu	Glu	Leu	Asn	Ala	Ile	Asn	Thr	Thr	Val	Ser	Gln	Asp
-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----

09868605 "091201"

COMBINED DECLARATION AND POWER OF ATTORNEY FOR PATENT APPLICATION

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled IMPROVEMENT OF TOLERANCE TO A XENOGRRAFT, the specification of which

- ☒ is attached hereto.
- ☐ was filed on _____ as United States Application No. _____.
- ☒ was filed on 17 December 1999 as International Application No. PCT/GB99/04200.
- ☐ and was amended on _____ (if applicable).
- ☐ with amendments through _____ (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, § 1.56. If this is a continuation-in-part application filed under the conditions specified in 35 U.S.C. § 120 which discloses and claims subject matter in addition to that disclosed in the prior copending application, I further acknowledge the duty to disclose material information as defined in 37 C.F.R. § 1.56 which occurred between the filing date of the prior application and the national or PCT international filing date of the continuation-in-part application.

I hereby claim foreign priority benefits under Title 35, United States Code, § 119(a)-(d) of any foreign application(s) for patent or inventor's certificate or of any PCT International application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application(s) for patent or inventor's certificate or any PCT International application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) on which priority is claimed:

Prior Foreign Application(s)			Priority Claimed	
<u>9827921.9</u>	<u>United Kingdom</u>	<u>19 December 1998</u>	<input checked="" type="checkbox"/>	<input type="checkbox"/>
<u>9925015.1</u>	<u>United Kingdom</u>	<u>23 October 1999</u>	<input checked="" type="checkbox"/>	<input type="checkbox"/>
(Number)	(Country)	(Day/Month/Year Filed)	Yes	No

I hereby claim the benefit under Title 35, United States Code, § 119(e) of any United States provisional application(s) listed below:

_____	_____
Application Number	Filing Date

0985605-094201

I hereby claim the benefit under Title 35, United States Code, § 120 of any United States application(s) or § 365(c) of any PCT International application(s) designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT International application in the manner provided by the first paragraph of Title 35, United States Code, § 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, § 1.56(a) which occurred between the filing date of the prior application and the national or PCT International filing date of this application:

PCT/GB99/04200
(Application No.)

17 December 1999
(Filing Date)

Pending
(Status: patented,
Pending, abandoned)

The undersigned hereby authorizes the U.S. attorney or agent named herein to accept and follow instructions from _____ as to any action to be taken in the Patent and Trademark Office regarding this application without direct communication between the U.S. attorney or agent and the undersigned. In the event of a change in the persons from whom instructions may be taken, the U.S. attorney or agent named herein will be so notified by the undersigned.

I hereby appoint the practitioners associated with the customer number provided below to prosecute this application, to file a corresponding international application, and to transact all business in the Patent and Trademark Office connected therewith:

Customer Number



24197
KSCLW

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JONCUS, Stephen J.	<u>44,809</u>	SIEGEL, Susan Alpert	<u>43,121</u>
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KLARQUIST, Kenneth S.	<u>16,445</u>	STEPHENS Jr., Donald L.	<u>34,022</u>
KLITZKE II, Ramon A.	<u>30,188</u>	STUART, John W.	<u>24,540</u>
LEIGH, James S.	<u>20,434</u>	VANDENBERG, John D.	<u>31,312</u>
MAURER, Gregory L.	<u>43,781</u>	WHINSTON, Arthur L.	<u>19,155</u>
NOONAN, William D.	<u>30,878</u>	WIGHT, Stephen A.	<u>37,759</u>
ORR, David E.	<u>44,988</u>	WINN, Garth A.	<u>33,220</u>

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I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under § 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

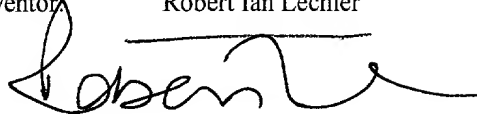
0986605-54103

28

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1-00


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Full Name of Third Inventor:

3-00

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A. Dorling

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Date

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09555505-091201

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Lys Tyr Leu Gly Arg Thr Ser Phe Asp Arg Asn Asn Trp Thr Leu Arg
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 Gln Gly His Pro Lys Pro Lys Lys Met Tyr Phe Leu Ile Thr Asn Ser
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 Glu Leu Phe Ser Ile Ser Asn Ser Leu Ser Leu Ser Phe Pro Asp Gly
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 Val Trp His Met Thr Val Val Cys Val Leu Glu Thr Glu Ser Met Lys
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 Ile Ser Ser Lys Pro Leu Asn Phe Thr Gln Glu Phe Pro Ser Pro Gln
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 Thr Tyr Trp Lys Glu Ile Thr Ala Ser Val Thr Val Ala Leu Leu Leu
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Arg His Cys Glu Pro Asn Gln Gly Leu Arg Val Lys Lys Glu Gly Thr
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Ser Lys Asp Cys Glu Ala Cys Ala Gln His Thr Pro Cys Ile Pro Gly
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Phe Gly Val Met Glu Met Ala Thr Glu Thr Thr Asp Thr Val Cys His
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Cys Tyr Pro Trp Thr Ser Cys Glu Asp Lys Asn Leu Glu Val Leu Gln
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195 200 205

Phe Gly Val Phe Leu Tyr Ile Lys Lys Val Val Lys Lys Pro Lys Asp
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Val

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<210> 14
 <211> 330
 <212> PRT
 <213> Porcus spp

<400> 14
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 Pro Cys His Phe Thr Asn Ser Gln Asn Leu Ser Leu Asp Glu Leu Val
 35 40 45
 Ile Phe Trp Gln Asp Gln Asp Asn Leu Val Leu Tyr Glu Leu Tyr Arg
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 Gly Gln Glu Lys Pro His Asn Val Asn Ser Lys Tyr Met Gly Arg Thr
 65 70 75 80
 Ser Phe Asp Gln Ala Thr Trp Thr Leu Arg Leu His Asn Val Gln Ile
 85 90 95
 Lys Asp Lys Gly Ser Tyr Gln Cys Phe Ile His His Lys Gly Pro His
 100 105 110
 Gly Leu Val Pro Ile His Gln Met Ser Ser Asp Leu Ser Leu Leu Ala
 115 120 125

096655 "091201

Asn Phe Ser Gln Pro Glu Ile Asn Leu Leu Thr Asn His Thr Glu Asn
 130 135 140
 Ser Val Ile Asn Leu Thr Cys Ser Ser Thr Gln Gly Tyr Pro Glu Pro
 145 150 155 160
 Gln Arg Met Tyr Met Leu Leu Asn Thr Lys Asn Ser Thr Thr Glu His
 165 170 175
 Asp Ala Asp Met Lys Lys Ser Gln Asn Asn Ile Thr Glu Leu Tyr Asn
 180 185 190
 Val Ser Ile Arg Val Ser Leu Pro Ile Pro Pro Glu Thr Asn Val Ser
 195 200 205
 Ile Val Cys Val Leu Gln Leu Glu Pro Ser Lys Thr Leu Leu Phe Ser
 210 215 220
 Leu Pro Cys Asn Ile Asp Ala Lys Pro Pro Val Gln Pro Pro Val Pro
 225 230 235 240
 Asp His Ile Leu Trp Ile Ala Ala Leu Leu Val Thr Val Val Val Val
 245 250 255
 Cys Gly Met Val Ser Phe Val Thr Leu Arg Lys Arg Lys Lys Lys Gln
 260 265 270
 Pro Gly Pro Ser Asn Glu Cys Gly Glu Thr Ile Lys Met Asn Arg Lys
 275 280 285
 Ala Ser Glu Gln Thr Lys Asn Arg Ala Glu Val His Glu Arg Ser Asp
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 Ser Thr Thr Asp Phe Leu Lys Ser Lys Leu
 325 330

<210> 15
 <211> 837
 <212> DNA
 <213> Porcus

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 gatctggagg attttcccga ctccaccgct ccggtgcagg agaccttaca ttggtgccag 780
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05863505-091201

<210> 16
 <211> 278
 <212> PRT
 <213> Porcus

<400> 16

Met Val Arg Leu Pro Leu Gln Cys Leu Leu Trp Gly Cys Phe Leu Thr
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Ala Val His Pro Glu Pro Pro Thr Ser Cys Lys Glu Asn Gln Tyr Pro
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Thr Asn Ser Arg Cys Cys Asn Leu Cys Pro Pro Gly Gln Lys Leu Val
 35 40 45

Asn His Cys Thr Glu Val Thr Glu Thr Glu Cys Leu Pro Cys Ser Ser
 50 55 60

Ser Glu Phe Leu Ala Thr Trp Asn Arg Glu Lys His Cys His Gln His
 65 70 75 80

Lys Tyr Cys Asp Pro Asn Leu Gly Leu Gln Val Gln Arg Glu Gly Thr
 85 90 95

Ser Lys Thr Asp Thr Thr Cys Val Cys Ser Glu Gly His His Cys Thr
 100 105 110

Asn Ser Ala Cys Glu Ser Cys Thr Leu His Ser Leu Cys Phe Pro Gly
 115 120 125

Leu Gly Val Lys Gln Met Ala Thr Glu Val Ser Asp Thr Ile Cys Glu
 130 135 140

Pro Cys Pro Val Gly Phe Phe Ser Asn Val Ser Ser Ala Ser Glu Lys
 145 150 155 160

Cys Gln Pro Trp Thr Ser Cys Glu Ser Lys Gly Leu Val Glu Gln Arg
 165 170 175

Ala Gly Thr Asn Lys Thr Asp Val Val Cys Gly Phe Gln Ser Arg Met
 180 185 190

Arg Ala Leu Val Val Ile Pro Ile Thr Leu Gly Ile Leu Phe Ala Val
 195 200 205

Leu Leu Val Phe Leu Cys Ile Arg Lys Val Thr Lys Glu Gln Glu Thr
 210 215 220

Lys Ala Leu His Pro Lys Thr Glu Arg Gln Asp Pro Val Glu Thr Ile
 225 230 235 240

Asp Leu Glu Asp Phe Pro Asp Ser Thr Ala Pro Val Gln Glu Thr Leu
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His Trp Cys Gln Pro Val Thr Gln Glu Asp Gly Lys Glu Ser Arg Ile
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Ser Val Gln Glu Arg Gln
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0966505-091201

<210> 17
 <211> 534
 <212> PRT
 <213> Porcus

<400> 17

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Glu	Ser	Ser	Leu	Ser	Phe	Ser	Trp	Arg	Thr	Gln	Ile	Asp	Ser	Pro	Leu
	50					55					60				
Asn	Gly	Lys	Val	Lys	Thr	Asn	Gly	Thr	Arg	Ser	Thr	Leu	Val	Met	Asn
65					70					75					80
Pro	Val	Ser	Phe	Glu	Asn	Glu	His	Ser	Tyr	Leu	Cys	Thr	Val	Ser	Cys
				85					90					95	
Gly	Asn	Leu	Lys	Gly	Glu	Arg	Gly	Ile	Gln	Val	Glu	Ile	Tyr	Ser	Phe
			100					105					110		
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		115					120					125			
Pro	Val	Thr	Val	Arg	Cys	Leu	Val	Pro	Asp	Val	Tyr	Pro	Val	Glu	Lys
	130					135					140				
Leu	Glu	Ile	Glu	Leu	Leu	Lys	Asp	Asn	His	Ser	Met	Val	Ser	Gln	Asn
145					150					155					160
Phe	Leu	Glu	Leu	Ile	Asp	Ile	Lys	Ser	Lys	Glu	Thr	Lys	Ser	Leu	Glu
				165					170					175	
Phe	Thr	Phe	Thr	Pro	Thr	Glu	Glu	Asp	Ile	Gly	Lys	Ala	Ile	Val	Cys
			180					185						190	
Gln	Ala	Thr	Leu	Ile	Ile	Asp	Gly	Gln	Pro	Ser	Val	Lys	Thr	Thr	Pro
		195					200					205			
Glu	Lys	Met	Gln	Val	Tyr	Ile	Ser	Pro	Lys	Asp	Pro	Val	Ile	Ser	Val
	210					215					220				
Asn	Pro	Ser	Thr	Ser	Leu	Gln	Glu	Gly	Asp	Ser	Met	Met	Met	Thr	Cys
225					230					235					240
Thr	Ser	Glu	Gly	Leu	Pro	Ala	Pro	Gln	Ile	Ser	Trp	Ser	Lys	Lys	Leu
				245					250					255	
Asp	Asn	Gly	Asp	Gln	Gln	Leu	Leu	Ser	Gly	Asn	Ala	Thr	Leu	Thr	Leu
			260					265					270		
Ile	Ala	Met	Arg	Met	Glu	Asp	Ser	Gly	Ile	Tyr	Val	Cys	Glu	Gly	Val
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090505 "091201"


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<210> 19
 <211> 269
 <212> PRT
 <213> Vacca spp

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<400> 19
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Val Asn Ser Leu Cys Cys Asp Leu Cys Pro Pro Gly Gln Lys Leu Val
      35              40              45

Asn Asp Cys Thr Glu Val Ser Lys Thr Glu Cys Gln Ser Cys Gly Lys
      50              55              60

Gly Glu Phe Leu Ser Thr Trp Asn Arg Glu Lys Tyr Cys His Glu His
      65              70              75              80

Arg Tyr Cys Asn Pro Asn Leu Gly Leu Arg Ile Gln Ser Glu Gly Thr
      85              90              95

Leu Asn Thr Asp Thr Ile Cys Val Cys Val Glu Gly Gln His Cys Thr
      100             105             110

Ser His Thr Cys Glu Ser Cys Thr Pro His Ser Leu Cys Leu Pro Gly
      115             120             125

Phe Gly Val Lys Gln Ile Ala Thr Gly Leu Leu Asp Thr Val Cys Glu
      130             135             140

Pro Cys Pro Leu Gly Phe Phe Ser Asn Val Ser Ser Ala Phe Glu Lys
      145             150             155             160

Cys His Arg Trp Thr Ser Cys Glu Arg Lys Gly Leu Val Glu Gln His
      165             170             175

Val Gly Thr Asn Lys Thr Asp Val Val Cys Gly Phe Gln Ser Arg Met
      180             185             190

Arg Thr Leu Val Val Ile Pro Val Thr Met Gly Val Leu Phe Ala Val
      195             200             205

Leu Leu Val Ser Ala Cys Ile Arg Asn Ile Thr Lys Lys Arg Gln Leu
      210             215             220

Arg Pro Cys Thr Leu Trp Leu Lys Gly Arg Ile Pro Trp Arg Arg Leu
      225             230             235             240

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09554505.091204

Ile Arg Arg Ile Phe Pro Ala Pro Thr Arg Leu Ser Gly Ala Arg Asp
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<210> 20
 <211> 867
 <212> DNA
 <213> Vacca spp

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 caaactcgca tctactggca aaaggagaag aaaatggtgc tgactatgat gtctggggac 240
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 gataacctgc tcccatcctg ggccattacc ttaatctcag taaatggaat ttttgtgata 780
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<210> 21
 <211> 35
 <212> DNA
 <213> Porcus spp

<400> 21
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<210> 22
 <211> 34
 <212> DNA
 <213> Porcus

<400> 22
 gcatgtcgac ttaaaaatct gtagtactgt tgtc 34

<210> 23
 <211> 17
 <212> DNA
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<400> 23
 agaccgtctt ccttttag 17

<210> 24
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CCDS:69120.1

<400> 24
ttggatcctc catgttatcc c 21

<210> 25
<211> 12
<212> DNA
<213> Porcus

<400> 25
agcatctgaa gc 12

<210> 26
<211> 22
<212> DNA
<213> Porcus spp

<400> 26
atggatcctc cattttccaa cc 22

<210> 27
<211> 18
<212> DNA
<213> Porcus spp

<400> 27
ttgtcgacat ctactggc 18

<210> 28
<211> 58
<212> DNA
<213> Porcus spp

<400> 28
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<210> 29
<211> 29
<212> DNA
<213> Porcus spp

<400> 29
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<210> 30
<211> 27
<212> PRT
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<220>
<223> Description of Artificial Sequence: Porcus spp/ovalbumen
chimeric peptide

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094655-091201

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<210> 31
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 <212> PRT
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<400> 31
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 20 25

<210> 32
 <211> 30
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 <213> Artificial Sequence

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 <223> Description of Artificial Sequence: Porcus spp/ovalbumen
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<400> 32
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Arg Lys Gly Pro His Gly Leu Val Pro Ile His Gln Met Ser
 20 25 30

<210> 33
 <211> 26
 <212> PRT
 <213> Artificial Sequence

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 <223> Description of Artificial Sequence: Porcus spp/ovalbumen
 chimeric peptide

<400> 33
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Arg Gly Leu Val Pro Ile His Gln Met Ser
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<210> 34
 <211> 28
 <212> PRT
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090605-091204

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<223> Description of Artificial Sequence: Porcus spp/ovalbumen
chimeric peptide

<400> 34

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<210> 35

<211> 29

<212> PRT

<213> Artificial Sequence

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<223> Description of Artificial Sequence: Porcus spp/ovalbumen
chimeric peptide

<400> 35

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<210> 36

<211> 29

<212> PRT

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chimeric peptide

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1 5 10 15

Arg Lys Ser Gln Ala Tyr Phe Asn Glu Thr Gly Glu Leu
20 25

<210> 37

<211> 29

<212> PRT

<213> Artificial Sequence

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Arg Ala Ser Leu Lys Ser Gln Ala Tyr Phe Asn Glu Thr

090605-091201

<210> 38
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 <212> PRT
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 chimeric peptide

<400> 38
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 20 25 30

<210> 39
 <211> 17
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Description of Artificial Sequence: Porcus spp/ovalbumen
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<400> 39
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Arg

090605-091201